

How to Improve the Bioreceptivity of Concrete Infrastructure Used in Marine Ecosystems? Literature Review for Mechanisms, Key Factors, and Colonization Effects

Mahmoud Hayek, Marie Salgues, Jean-Claude Souche, Klaartje de Weerdt,

Sylvain Pioch

► To cite this version:

Mahmoud Hayek, Marie Salgues, Jean-Claude Souche, Klaartje de Weerdt, Sylvain Pioch. How to Improve the Bioreceptivity of Concrete Infrastructure Used in Marine Ecosystems? Literature Review for Mechanisms, Key Factors, and Colonization Effects. Journal of Coastal Research, 2023, 39 (3), pp.553-568. 10.2112/JCOASTRES-D-21-00158.1. hal-04051748

HAL Id: hal-04051748 https://imt-mines-ales.hal.science/hal-04051748v1

Submitted on 3 Apr 2023 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

How To Improve The Bioreceptivity of Concrete Infrastructure Used in 1 Marine Ecosystems? Literature Review for Mechanisms, Key Factors and 2 **Colonization Effects** 3 Mahmoud Hayek[†], Marie Salgues[†], Jean-Claude Souche[†], Klaartje De Weerdt[‡], Sylvain 4 **Pioch[§]** 5 [†] LMGC, IMT Mines Alès 6 7 Univ Montpellier, CNRS Alès, France 8 [‡] Department of Structural Engineering 9 Norwegian University of Science and Technology NTNU 10 Trondheim, Norway 11 [§]LAGAM 12 University Paul Valéry Montpellier 3 13 Montpellier, France 14 sylvain.pioch@univ-montp3.fr 15 LRH: Hayek, Salgues, Souche, De Weerdt, Pioch 16 **RRH:** Bioreceptivity of marine concrete 17

18

ABSTRACT

19 With 39,400 km² of coastal and marine areas artificialized and an increasing demand due to the growing global population – 9 billion by 2050 – we need to find ways to mitigate futures construc-20 tions impacts on biodiversity. In this study, we explore how civil engineering can take further tech-21 nical measures to enhance marine biodiversity, in a real and valuable "win-win" strategy. The 22 global aim is to integrate eco-engineering practices within coastal projects and include ecological 23 targets (e.g. the diversity and speed of biological colonization) early, at the project design stage, 24 25 with the same level efforts for technical, social and economic studies. Concrete is the most useful material for coastal infrastructures constructions, therefore enhance its positive impact on coloni-26 zation is by far one of the key-point for developers and coastal managers. To this end, we've first 27 reviewed the latest research regarding the bioreceptivity of concrete, focusing on the characteristics 28 of the marine environment that affect the colonization of concrete and the organisms involved. 29 From this base of publications, we've updated the intrinsic and environmental parameters that can 30 31 influence the intrinsic and the extrinsic bioreceptivity of concrete and operate the link with the mechanisms leading to the colonization of concrete and biofilm formation, which wasn't done be-32 fore. Based on the persistence of their significant effect (after 78 days of immersion in seawater), 33 the intrinsic parameters that support greater biocolonization are classified from more to less effec-34 35 tive in the following order; surface roughness (190%) > chemical composition (slag cement instead Portland cement) (136 %) > chemical composition (presence of formwork oil) (106 %). We've 36

- finally analyzed both the ecological effect and the positive and negative effects of biofilm for-mation on the durability of concrete, in the scope to provide clear and operational results for future
- 39 concrete coastal construction implementation for decision maker.
- 40

ADDITIONAL INDEX WORDS

- 41 *Eco-engineering practices, marine biodiversity, concrete, colonization, eco-design, bioreceptivity,*
- 42 *biofilm, durability.*
- 43
- 44

INTRODUCTION

45

46 The global ecological crisis is the concomitant with the Anthropocene, a new geological era in which humans have become the central actors on the planet (Crutzen, 2006; Shukla et al., 2021; 47 Valipour, Bateni, and Jun, 2021). The recent report (2019) of the Intergovernmental Science-Policy 48 Platform on Biodiversity and Ecosystem Services (IPBES) presents a devastating assessment of 49 the impact on biodiversity since the start of the industrial era two centuries ago: in terms of coastal 50 and marine ecosystems, more than 85% of wetlands have been destroyed, and human activities 51 threaten 66% of the oceans. The causes leading to this situation are today well known and include 52 (in order of impact): (1) artificialization and land use, (2) overexploitation of resources (fishing, 53 54 forestry, etc.), (3) climate change, (4) pollution (plastics, chemical residues, etc.) and (5) invasive species (Ibid). 55

Therefore, reducing artificialization and land use is critical to reduce biodiversity loss and preserve 56 the future of humanity. Yet predictions indicate a clear acceleration of urbanization around the 57 world, on land and at sea. Between 2019 and 2040, more than US\$60,000 billion is forecast to be 58 59 invested in infrastructure in the 56 countries representing 88% of global GDP (Global Infrastructure Hub 2017–2018). More than 1.2 million km² will be urbanized by 2030 – just 8 years 60 61 from now - representing an increase of more than 185% compared to the period from 1970 to 2000 62 (Seto et al., 2012). By 2050, 3 to 4.7 million km² of roads will be constructed, which represents an 63 increase of more than 25% compared to the current annual rate (Meijer et al., 2018). This process will be particularly significant along coasts, where eight of the ten largest megalopolises in the 64 world are located (such as Lagos, Tokyo, Jakarta and New York). Around the world, the population 65 living within 100 km of a coast is estimated to approach 7.5 billion people by 2050 (de Fraiture et 66 al., 2007), leading to a need for increased coastal development and potentially disturbing essential 67 coastal habitats (Weilgart, 2007). Global marine infrastructure extended over approximately 68 32,000 km² in 2018 (Bugnot et al., 2020) and is expected to reach 39,400 km² by 2028: a surface 69 area equivalent to the country of Bhutan. The area of seascapes impacted by these structures was 70 estimated to be between 1 and 3.4 million km² in 2018, which is expected to increase 50–70% by 71 2028 (Ibid). The phenomenon of 'coastal squeeze', a term introduced by Doody (2004) to refer to 72 73 the threat to the existence of coastal habitats caused by the compound impacts of sea-level rise and human activities, will not abate under current development conditions. Most of these structures are 74 made from concrete, the most durable materials for the aggressive marine environment. Table 1 75 summarize the types of concrete structures found in coastal areas. 76

From the 1970s through the 1990s, coastal ecology literature dealing with biodiversity focused 77 mainly on artificial substrates use to promote restoration, studied from colonization or settlement 78 phenomena problematics (Relini et al., 1998; Zobel, 1974). However, since the 2000s, hard 79 artificial infrastructure made with concrete, has been implicated as a major risk factor for local and 80 native biodiversity, by introducing invasive species with ubiquitous and rapid growing 81 characteristics, in comparison with surrounding natural rocky area (Bulleri and Airoldi, 2005; 82 Bulleri and Chapman, 2010, 2004; Jackson et al., 2001). In contrast, several studies about 83 ecological design of coastal infrastructure published since 2010 has clearly demonstrated that it is 84 85 feasible to enhance biodiversity through changes in concrete chemical composition, roughness,

- surface treatment, or inclusion of variously sized pits or holes (Chapman and Underwood, 2011;
- 87 Evans, Bax and Bernal, 2016; Evans, Bax and Smith, 2018; Martins et al., 2010; Phan, van Thiel
- de Vries and Stive, 2015; Souche et al., 2016). In addition, other studies also has indicated that
- 89 physical alteration and damage to artificial structures due to natural processes (e.g. weathering,
- 90 wave action, chemical erosion) could be reduced by the positive protective role of marine
- organisms (Coombes et al., 2017, 2013; Gowell, Coombes and Viles, 2015; La Marca et al., 2014;
- 92 Naylor and Coombes, 2015; Perkol-Finkel and Sella, 2014).
- 93 Table 1. Types of structures in coastal areas, their functions and the materials used **Types of** Socio-technical objectives **Materials** structure Coastal defense structures (protection of coastlines and management of wave action effects) **Offshore** Coastal defense structure built near the coast to Rock fill breakwater protect it from the actions of the sea by reducing Concrete blocks wave-action intensity Groin Coastal defense structure to protect the coast Rock fill from erosion and sediment transport Harbor Structure to protect a harbor area from wave Rock fill and blocks of concrete action and currents breakwater, (blocks, tetrapodes, accropodes) jetty Harbor structures Reinforced or prestressed Finger pier Mooring structure for loading and unloading concrete apron on caisson or boats gabion-type piles **Boom wharf** Structure on piles making up a finger pier (wharf Reinforced concrete apron on perpendicular to the coast) metal or reinforced concrete piles Continuous open-platform structure for Reinforced concrete caissons, Quay mooring, loading and unloading boats metal sheet-pile gabions, curtain walls and screens in metal or reinforced concrete Metal or reinforced concrete, held Floating dock Floating structure enabling docking and mooring of boats in a harbor in place by piles or reinforced concrete moorings Floating Reinforced concrete, held in place Floating structure to protect a harbor from wave breakwater by piles or reinforced concrete action moorings Lock / gate Structure enabling boats to change level and/or Infrastructure in reinforced to keep a dock filled with water concrete, equipment in steel

Dry dock	Waterproof basin that can be drained or filled with water to enable maintenance and repair of boats/caissons	Unreinforced concrete, reinforced concrete / reinforced concrete, prestressed concrete
Slipway	Slope enabling a boat to be launched or landed for maintenance	Reinforced concrete
Self-elevating platform, jack- up barge	Floating deck using a wet dock to raise boats out of the water for maintenance or refitting	Infrastructure in reinforced concrete, equipment in steel
Open-sea struct	ures	
Offshore structure	Structure out as sea such as an oil rig or wind turbine	Reinforced concrete and steel
Anchorage, ballast	Underwater structure for anchoring boats or floating infrastructure	Reinforced concrete
Artificial reef	Underwater structure for protecting or hosting fauna and flora or for leisure activities	Reinforced concrete, metal, diverse materials

Harbor structures for maintenance and services

94

Pragmatically, most current coastal concrete infrastructure cannot be readily removed. But there is 95 now increasing research efforts into innovative ways that future new infrastructures can be 96 designed and built to meet engineering requirements, while also increasing its value as an 97 ecological habitat for marine life (Dafforn et al., 2015; Firth et al., 2014; Lacroix and Pioch, 2011; 98 Naylor and Coombes, 2015; Patranella et al., 2017; Perkol-Finkel and Sella, 2014; Phan, van Thiel 99 de Vries and Stive, 2015; Pioch and Souche, 2021). This "win-win" approach, should ensure 100 101 benefits both for human society and nature (Chapman and Underwood, 2011; Ferrario et al., 2014; Perkol-Finkel et al., 2018; Pioch et al., 2018, 2011; Temmerman et al., 2013). This paradigm 102 support the idea that we have to construct structures with a minimal ecological impact on the 103 104 existing environment, maximizing their potential for colonization, and restore original habitats or create new habitats adapted for local species ecological requirements (Browne and Chapman, 105 2011; Souche et al., 2018). In this way, combining engineering techniques with ecological 106 107 understanding can provide, with a cost-effective ways, the maintenance or development of native biodiversity (Airoldi et al., 2020; Martins et al., 2010; Pioch et al., 2018; Scheres and Schüttrumpf, 108 109 2019; Souche et al., 2019).

Artificial marine structures modify, positively or negatively, the natural environment (Azevedo *et al.*, 2006) to become part of a new ecosystem (Giraudel, Garcia and Ledoux, 2014). Then to avoid high negatives impacts, as we have seen, the ecological mechanism of settlement on concrete needs to be assessed before their implementation (Baine, 2001; Bulleri and Chapman, 2010). Impacts parameters are divided in five main fields: localization for the immersion zone, implantation (Giraudel, Garcia and Ledoux, 2014), seasons and immersion time (Firth *et al.*, 2014; Vaz-Pinto *et al.*, 2014), design of the structure (Baine, 2001; Pioch *et al.*, 2011), and the type of materials used (Anderson and Underwood, 1994; Fitzhardinge and Bailey-Brock, 1989; Lee *et al.*, 2008). There
are also interdependent relationships that change over time between the type of structure, exposure
conditions, substrate, seasonality and colonization by living organisms (Anderson and Underwood,
1994; C. Choi *et al.*, 2006; Somsueb, Ohno and Kimura, 2001).

121 The way to associate civil engineering and ecological consideration is now a common subject of research with eco-engineering, and specifically within the construction field the eco-design 122 strategies. Recent research applications, have demonstrated that it's possible to integrate 123 environmental objectives in the design of functional concrete marine infrastructures (Bulleri and 124 Chapman, 2010; Lacroix and Pioch, 2011; Pioch et al., 2011; Pioch and Souche, 2021). Thus, 125 understanding the interactions between microorganisms and cementitious materials is a crucial step 126 toward constructing more durable, safer and more ecological structures (Lors, Feugeas and 127 Tribollet, 2019; Ottelé et al., 2011; Perini and Ottelé, 2014; Perkol-Finkel and Sella, 2014). As the 128 colonization of cementitious material by microorganisms is an essential step for the adhesion of 129 other organisms (Mieszkin, Callow and Callow, 2013; Rajitha et al., 2020; Salta et al., 2013), the 130 aim is to favor this colonization in order to reduce the ecological impact of the structure and 131 consequently increase biodiversity, without impacting the structure's strength and durability. Then, 132 additional studies are necessary to determine the different factors that can influence the 133 colonization of cementitious materials in the marine environment, but the main parameters seem 134 to be the water pH, the materials chemical composition and surface roughness or topography 135 (Grosseau et al., 2015; Hayek et al., 2020a, 2021; McManus et al., 2018; Miller et al., 2009, 2012). 136

137 The objective of this study was to identify key parameters to take into account to promote the 138 bioreceptivity of concrete structures in coastal and marine environments to identify key parameters 139 to take into account in order to promote the bioreceptivity of concrete structures.

140 Scope and research significance

The interaction of cement-based materials with microorganisms has been widely studied in 141 building and sewer network. To the best of our knowledge, in the marine environment, the present 142 143 literature review is the first one in the field of concrete bioreceptivity and concrete biocolonization by marine bacteria. In this review, a significant amount of research has been performed on the first 144 stage of colonization of concrete in seawater with significant research on this topic including i) the 145 early colonization process and the detailed mechanisms involved in the biofilm formation on the 146 surface of marine concrete ii) the colonizing organisms of cement-based materials and iii) the 147 physicochemical factors influencing the colonization of cement-based materials in marine 148 149 environments.

- 150 Afterward, to achieve the purpose of this paper, i) some of the most significant quantitative results
- 151 about the bioreceptivity of concrete in the marine environment are presented ii) the ecological
- impact and the effect of biological colonization on the durability of concrete are discussed and iii)
- the perspectives to design a more bioreceptive concrete (green concrete) are mentioned.
- 154 Web of science and google scholar are the two databases used in this study for collecting the 155 relevant materials (211 studies) from the literature. The keywords used were: Biofilm formation,

Concrete biocolonization, Ecological impact, Eco-engineering *practices* Artificial reef, Marine
 environment, Concrete Durability, Bioreceptivity and marine biodiversity.

METHODS: BIOLOGICAL FACTORS AFFECTING THE

COLONIZATION OF CONCRETE IN THE MARINE

ENVIRONMENT

158

159

160

161

162 Colonization and the development of biofilm on surface concrete

From the first hours of the immersion of an object in seawater, a cement-based material is covered with a biofilm consisting of one or more microbial communities (Dang and Lovell, 2015; Dobretsov, Teplitski and Paul, 2009). By definition, the biofilm is considered to be a sessile microbial community irreversibly attached to a substrate or an interface and included in a coating of extracellular polymeric substances (EPS). Biofilm cells have modified phenotypes compared to plankton cells that float freely in the environment (Donlan and Costerton, 2002).

The biofilm is the first stage in colonization, which enables the establishment of other marine organisms via the EPS synthesized on the surface (Hinsa and O'Toole, 2006; Pasmore and Costerton, 2003; Wood *et al.*, 2016). Bacteria (bacterial biofilm) are the first colonizers, which facilitate the adhesion of other micro- and macro-organisms such as fungi, microalgae, macroalgae and invertebrates (Briand *et al.*, 2018; Brian-Jaisson *et al.*, 2014; Dobretsov and Rittschof, 2020; Salta *et al.*, 2013) (Figure 1).

After immersion, an induction phase allows the surface to be conditioned (through the formation 175 of the conditioning film), resulting in exponential accumulation, characterized by the adhesion and 176 growth of bacteria and other microorganisms on the surface. The length of the induction phase 177 depends on both the environmental parameters and the intrinsic parameters of the host material 178 (Hayek et al., 2019; Karunasagar et al., 1996; Katsikogianni and Missirlis, 2004). A submerged 179 solid object adsorbs organic and inorganic molecules and ions, leading to a modification of the 180 physicochemical properties of the surface, thus forming a nutrient-rich micro-niche (on a small 181 scale in comparison to the material) that favors the adhesion of bacterial cells. In this way, surface 182 conditioning impacts the adhesion of bacterial cells (Loeb and Neihof, 1975), which can be 183 stimulated or inhibited by the nature of the adsorbed molecules. This is why the conditioning of 184 the surface is considered a key step in biofilm formation (Herrmann *et al.*, 1988; Tang *et al.*, 2006). 185

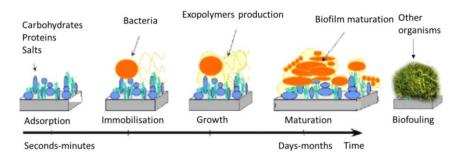




Figure 1. Biofouling in the marine environment (Ogunola and Onada, 2016).

188 Therefore, the main stages of biofilm formation are (i) reversible and irreversible adhesion of 189 bacterial cells to the surface of the cement-based material; (ii) growth and maturing of the biofilm 190 with the secretion of EPS; and (iii) partial detachment of the biofilm and the dispersion of bacterial

191 cells (Hinsa and O'Toole, 2006; O'Toole, Kaplan and Kolter, 2000).

192 The adhesion of bacteria to a surface first requires that they move toward the surface to be 193 colonized. This movement can be due to environmental phenomena (gravitational and 194 hydrodynamic forces in the surrounding fluid) and/or by appendages on the bacteria that assist their 195 movement, such as flagella or pili (Ha and O'Toole, 2015; Katsikogianni and Missirlis, 2004).

196 Once a bacterium is within a critical distance of the object (generally < 1 nm), low-intensity interactions take place between the bacterium and the surface, e.g. electrostatic forces, Van der 197 Waals forces and hydrophobic interactions (Carpentier and Cerf, 1993; Dunne, 2002). Depending 198 on these various interactions, the bacterium may either adhere or be repelled by the effect of 199 hydrodynamic actions. If the interactions are favorable, the bacterium adheres to the surface and 200 this step is then considered the reversible adhesion phase (An and Friedman, 1998; Berne et al., 201 2018; Busscher and Weerkamp, 1987; van Loosdrecht et al., 1989). The factors that influence this 202 phase concern interactions between the bacterial cells and the surface of the cement-based material: 203 (i) the specific characteristics of the bacteria, such as its cell surface charge, cell surface 204 hydrophobicity and the chemical composition of the cell wall; (ii) the intrinsic parameters of the 205 host surface, such as its physicochemical nature, polarity, porosity and roughness; and (iii) the 206 environmental conditions, such as pH level, temperature and flow conditions (Bunt et al., 1993; 207 Scheuerman et al., 1998; Speranza et al., 2004). According to Carpentier and Cerf (1993), host 208 surface porosity and roughness are believed to increase bacterial adhesion, while a negatively 209 charged surface may have a repulsive effect as most bacteria are negatively charged. 210

Adhesion subsequently becomes irreversible when strong covalent interactions are established between the bacteria and the surface of the cement-based material. The covalent interactions are formed by macromolecules on the bacteria surface and include polysaccharides, proteins or lipopolysaccharides (Berne *et al.*, 2018).

Once established on the object, the bacteria multiply and the EPS they secrete develop a three-215 dimensional structure to form colonies that cover all or part of the surface. This varies depending 216 on the bacteria present and (i) their capacity to communicate with each other, i.e. their cell-to-cell 217 communication, or quorum sensing, system (Branda et al., 2005; Dobretsov et al., 2011); (ii) the 218 intracellular concentration of cyclic di-GMP (a high intracellular concentration of c-di-GMP 219 promotes the expression of genes involved in biofilm formation) (Barraud et al., 2009; Chao et al., 220 2013; Dang and Lovell, 2015); (iii) environmental conditions, and (iv) the properties of the host 221 222 material, especially hydrophobic properties (O'Toole, Kaplan and Kolter, 2000).

223 A mature biofilm has a three-dimensional structure surrounded by a matrix made up of mineral

elements and EPS: proteins, polysaccharides, nucleic acids, lipids and other cellular components.

225 This matrix forms a barrier that protects biofilm microorganisms against external threats (Ha and

226 O'Toole, 2015; Soleimani *et al.*, 2013).

227 The last step in the development of the biofilm is the dispersion of cells in the environment after

- detaching from the surface. This dispersion is an essential part of the biofilm cycle as it enables the
- bacteria to colonize new surfaces and develop new biofilms (Kaplan, 2010). In general, biofilm
- dispersion mechanisms can be divided into two broad categories: (i) active dispersion based on
- mechanisms initiated by the bacteria; and (ii) passive dispersion due to external forces, such as shearing forces caused by hydrodynamic conditions (Lawrence *et al.*, 2002). Several factors are
- involved in the complex process of biofilm dispersion, such as environmental signals, quorum
- sensing systems, and the bacteria's production of extracellular enzymes that can degrade the matrix
- of biofilm components and facilitate dissociation (Boyd and Chakrabarty, 1994; Kaplan, 2010).

236 Colonizing organisms of cement-based materials

237 In the marine environment, there are many ubiquitous colonizing microorganisms likely to form a

biofilm on the surface of objects under suitable conditions. The main colonizing microorganisms
of cement-based materials are bacteria, fungi, cyanobacteria and algae (Gaylarde and Gaylarde,

240 2005).

241 Bacteria

Bacteria are medium-sized unicellular microorganisms that vary in size from 0.5 to 1.5 μ m (Yañez *et al.*, 2010) and develop on all types of surface materials; they are generally pioneers in the colonization process (Meng *et al.*, 2020; Romani *et al.*, 2019; Scatigno *et al.*, 2016).

The deterioration of cement-based materials by microbial induced corrosion has been known since 245 1945 following the work of Parker, who was the first to isolate from concrete samples a sulfuric 246 acid-producing bacterium species initially called *Thiobacillus concretivorus*, then subsequently 247 called Acidithiobacillus thiooxidans (Parker, 1945). This phenomenon has mainly been studied in 248 sanitation networks, where the sulfate- and nutrient-rich environment provides optimal conditions 249 for the development of microbial fauna belonging to the genus Thiobacillus (Yuan et al., 2015). 250 Thiobacilli are bacteria that oxidize sulfur compounds into sulfuric acid, a substance that is 251 particularly aggressive to concrete (De Belie, 2010). However, the biochemical processes leading 252 to the degradation of concrete under the influence of bacteria are essentially based on the synthesis 253 (bacterial metabolism) of acids such as sulfuric acid (H₂SO₄) and nitric acid (HNO₃) (Cwalina, 254 2008). These acids dissolve calcareous components, resulting in a loss of cohesion in the material 255 and the appearance of cracks or corrosion of the reinforcement bars in the concrete (Chatterjee and 256 Goyns, 2013; Poonguzhali et al., 2008). These effects are common in the sulfate- and ammoniac-257

258 rich environment of sanitation networks.

Despite the large number of concrete structures present in the waters of seas, rivers and lakes,
studies concerning the colonization of cementitious materials by bacteria in these environments
remain relatively rare (Chlayon, Iwanami et Chijiwa, 2020; Cooke *et al.*, 2020; Costa and
Appleton, 2001; Rica *et al.*, 2016; Scheres and Schüttrumpf, 2019; Sosa *et al.*, 2011; Souche *et al.*,
2016).

264 Algae and cyanobacteria

Microscopic algae (microalgae) and cyanobacteria are unicellular or multicellular species, generally between 0.5 μ m and 1 mm in size. These microorganisms are characterized by their capacity to perform oxygenic photosynthesis (Leclerc, 1975).¹ As algae and cyanobacteria are capable of generating their own organic matter from mineral elements, they are autotrophic organisms (Harper and Pendleton, 1993; Krumbein, 1979; Subashchandrabose et al., 2011).

According to the literature, green algae (Chlorophyta) and cyanobacteria (Cyanophyta) produce greenish, reddish or blackish stains on concrete, depending on the composition of the biofilm. In addition, the secretion of organic acids, resulting from the metabolism of these organisms, can lead to the dissolution of the calcium-rich phases of calcareous stones, concrete and mortar. In this way, algae and cyanobacteria can cause the alteration or degradation of cementitious materials (Escadeillas *et al.*, 2008, 2007).

- Cyanobacteria are prokaryotic microorganisms, whereas microalgae are eukaryotic. Microalgae 276 are divided into several phylogenetic groups, such as chlorophytes, rhodophytes and diatoms (Al-277 Handal and Wulff, 2008). Chlorophytes mainly develop in freshwater, such as rivers, and are the 278 largest group of green algae (Lueangthuwapranit, Sampantarak and Wongsai, 2011). Rhodophytes 279 are mainly macroscopic multicellular marine organisms and are known as red algae (Bengtson et 280 al., 2017). Diatoms are microscopic unicellular organisms characterized by a silica cell wall, called 281 a 'frustule', that surrounds them. They are abundant in the marine environment (Robert et al., 282 2004). 283
- The colonization of cement-based materials in situ by cyanobacteria and algae has been confirmed in various studies by measuring the quantity of chlorophyll and/or by the appearance of differentcolored stains on the samples tested (Silva and Naik, 2013; Souche *et al.*, 2016; Tran, 2011; Vishwakarma *et al.*, 2014). However, in most of these studies, the colonizing species were not identified and described, calling for further research to obtain a deeper understanding of the interactions between cement materials and microorganisms in the marine environment.

290 **Fungi**

291 Micro-fungi, whose size varies from 1 to 100 μ m, are chemotrophic eukaryotes. They are not 292 capable of photosynthesis to produce their own organic matter. Fungi require wet or moist 293 environments to develop, and their energy is provided by redox reactions. They are not very 294 sensitive to pH variation but develop more easily in acidic conditions.

Fungi are heterotrophs. They can only synthesize their cellular components from a source of organic carbon. Thus, fungal colonization of a cement-based material requires an accumulation of organic matter on the surface. In the case of successful colonization, fungi can then lead to the alteration or deterioration of the colonized material. Certain fungi produce pigments (for example, melanin) that cause dark stains on the cement-based material (Giannantonio et al., 2009). In addition, fungi produce a large quantity of organic acids (acetic, formic, oxalic and tartaric acid),

¹ Photosynthesis is the reaction enabling certain living organisms to synthesize organic matter from CO₂ and an electron donor under the effect of sunlight.

which are responsible for the dissolution of lime in mineral matrices and the chelation of calciumions, leading to the deterioration of the material.

The marine environment is considered to contain a large number of fungal organisms (Escadeillas *et al.*, 2007). However, only a few studies have focused on the colonization by fungi of cementbased materials in the marine environment. Submerged surfaces are often covered by one or more layers of algae and other protozoan organisms (Dürr and Thomason, 2009), which makes it difficult to identify and isolate sessile fungi colonizers in the marine environment without the use of advanced molecular biology techniques such as the extraction and sequencing of 18S rRNA.

309 <u>METHODS: PHYSICOCHEMICAL FACTORS INFLUENCING</u> 310 <u>THE COLONIZATION OF CEMENT-BASED MATERIALS IN</u> 311 <u>MARINE ENVIRONMENTS</u>

The biological colonization of a surface is governed by the interaction of three main factors: the type of material, the microorganisms and the environment (Lors, Feugeas and Tribollet, 2019). Then, two types of bioreceptivity are defined: (i) intrinsic bioreceptivity, linked only to the physicochemical properties of the concrete surface, such as porosity, roughness and chemical composition; and (ii) extrinsic bioreceptivity, related to the environment (Lors, Feugeas and Tribollet, 2019).

318 Intrinsic bioreceptivity (Material factors)

319 Chemical composition

Depending on its chemical composition, a material can contain substances that favor or inhibit the 320 development of biofilm (Ly et al., 2021). For example, the presence of certain heavy metals, in 321 sufficient quantities, can inhibit the growth and development of microorganisms (Dalod et al., 322 2014; Dubosc, Escadeillas and Blanc 2001). In a comparison of the bacterial colonization of 323 Portland cement (CEM I) and slag cement (CEM III) in seawater, CEM III mortar was shown to 324 be more bioreceptive than CEM I mortar (Hayek et al., 2020a, 2020b). Thus, the type of cement 325 influences the kinetics and the level of development of bacterial biofilm on a concrete surface. The 326 chemical composition of a concrete can also influence other important parameters, such as 327 roughness, porosity and pH (Veeger et al., 2021). 328

329 Surface roughness or topography

The roughness of the surface of a material is one of the main physical factors influencing its 330 bioreceptivity. Roughness consists of bumps and irregularities on the surface of the material. These 331 irregularities can act as anchoring sites and micro-refuges for microorganisms, thus favoring 332 colonization (Ammar et al., 2015). These micro-refuges protect microorganisms from the 333 hydrodynamic forces that tend to detach them during the highly precarious reversible adhesion 334 phase (Bellon-Fontaine et al., 2016). Kerr et al. (1999) had observed an increase in bacterial 335 adhesion on glass and acrylic samples immersed in natural seawater when roughness was raised 336 from 0.25 to 30 μ m. Teughels *et al.* (2006) had showed that the formation of dental plaque is 337 favored by a rough surface. Concerning cement-based materials, several studies have demonstrated 338

the effect of surface roughness on biofilm formation (Govin *et al.*, 2013; Hayek *et al.*, 2021; Manso *et al.*, 2014; Manso and Aguado, 2016; Miller *et al.*, 2009; Souche *et al.*, 2019; Tomaselli *et al.*, 2000; Tran *et al.*, 2014, 2012). These studies show that rough cement-based materials have higher bioreceptivity than smooth surfaces under both laboratory and natural conditions. To enhance colonization, the target is to obtain multiscale rugosity; it is possible to reach the required quality using special elastomeric formworks (Souche *et al.*, 2019).²

In addition to its effect on the adhesion of microorganisms, surface roughness also influences water 345 flow rates and the adsorption of organic matter (Dubosc, Escadeillas and Blanc 2001; El-Naggar 346 et al., 2010). This directly impacts the formation of the conditioning film during the induction 347 phase. Some studies nuance the effect of roughness on the bioreceptivity of a material, specifying 348 that this depends on the characteristics of the microorganisms involved, e.g. the type of 349 microorganism, geometry, size and shape (Whitman et al., 1998). For example, filamentous algae 350 such as *Klebsormidium flaccidum* attach to rough surfaces, whereas algae with thick cell walls 351 adhere to smooth surfaces (Lors, Feugeas and Tribollet, 2019; Rindi, Guiry and López-Bautista, 352 2008; Tran, 2011). 353

Total porosity to water

In the case of surfaces not immersed in water, bacteria preferentially colonize porous surfaces. This can mainly be explained by the influence of porosity on the absorption and retention of water by the material, a determining factor for the development of microorganisms (Nuhoglu *et al.*, 2006). Crevasses that are the same size as the microorganism concerned appear to be ideal. Smaller pits assist anchoring but do not protect from flows, while pits that are too large do not enable the optimum transfer of nutrients.

In the case of submerged concrete, it is difficult to determine the effect of porosity on bioreceptivity (Govin *et al.*, 2013; Tran *et al.*, 2014, 2012). This difficulty has been explained by the permanent saturation of samples tested under water and therefore the high humidity throughout the experiment.

365 Surface pH

The pH of the surface of the material can influence the type of microorganism and its capacity to adhere to the material (Shi and Zhu, 2009). Generally, each species has an optimum pH for growth and can develop within a more or less limited range around this value. Based on these pH ranges, microorganisms are classed as acidophilic (pH < 6), neutrophilic (6 < pH < 8) or basophilic (pH > 8).

The metabolic activities, growth and adhesion of microorganisms and the synthesis of EPS are influenced by the pH of the surface of the material. The reduction of the surface pH of concrete after immersion in seawater (from 13 to about 9) accelerates colonization of the surface and improves the bioreceptivity of the material (Dalod *et al.*, 2014; Hayek *et al.*, 2020b; Roberts *et al.*, 2002; Tran *et al.*, 2014; Vincke *et al.*, 2001). However, even concrete with a highly basic surface

² http://www.groupelib.com/les-moules-et-produits-en-elastomeres.html

could be colonized by extremophilic bacteria that can resist environmental conditions such asextreme temperature, pH or pressure.

378 Surface hydrophobicity

Surface hydrophobicity is one of the factors that affect the patterns and nature of bacterial 379 380 colonization of a surface, especially during the early stages of biofilm formation (Dang and Lovell, 2015; Scheuerman et al., 1998). Unsurprisingly, hydrophilic microorganisms attach more easily to 381 hydrophilic surfaces, whereas hydrophobic microorganisms prefer hydrophobic surfaces (Shi and 382 Zhu, 2009). As hydrophobicity and surface charge depend on the material's chemical composition 383 (Katsikogianni and Missirlis, 2004), this is one of the ways that composition influences the types 384 and diversity of colonizing microorganisms (Giannantonio et al., 2009; Guillitte, 1995; Miller et 385 al., 2012). 386

However, a hydrophobic surface exhibits low wettability and low surface energy, whereas a hydrophilic surface exhibits high wettability and high surface energy. Water droplets on a hydrophobic surface will 'bead up' while droplets on a hydrophilic surface will spread out evenly. The degree of hydrophobicity is determined by contact angle measurements between water droplet and surface; the surface is super-hydrophilic, hydrophilic, hydrophobic and superhydrophobic when the contact angle is respectively less than 10°, between 10° and 90°, between 90° and 150° and between 150° and 180° (Bixler and Bhushan, 2012).

394 Despite the importance of this parameter on marine biodiversity as well as on the colonization of submerged surfaces, studies that relate to the surface hydrophobicity of cementitious materials and 395 its effects on the biocolonization in the marine environment seem to be very rare. Hayek et al., 396 397 (2021) showed that the use of curing compound inhibits the biocolonization of concrete surface in seawater because of its effect on surface hydrophobicity (the cure compound forms a 398 superhydrophobic film on the concrete surface). This study is in agreement with the literature, 399 which has been proven that the use of a hydrophobic surface coating (low adhesive surface) is a 400 successful strategy to inhibit biofouling in the marine environment (Blainey and Marshall, 1991; 401 Huggett et al., 2009; Razavi et al., 2019). 402

403 Extrinsic bioreceptivity (Environmental factors)

404 Several studies have reported that the environment has a strong impact on the colonization of 405 cementitious materials in the marine context (Giraudel, Garcia and Ledoux, 2014; Lors, Feugeas 406 and Tribollet, 2019; Souche *et al.*, 2019). The medium influences the development and growth of 407 microorganisms as well as the aging of the material. Indeed, interactions between the environment 408 and microorganisms define the conditions in which microbial growth can occur (Figure 2).

Generally, the development of biofilm on the surface of a cement-based material is relatively slow as concrete has a high pH of about 13 (Codina, 2007; Manso and Aguado, 2017). This high alkalinity is unfavorable for biological colonization because few microorganisms can develop under such basic pH conditions. For that reason, an initial induction step that reduces the pH from 13 to 9.5 will favor the colonization of cement-based materials (Roberts *et al.*, 2002). This

414 modification of the surface pH arises from the adsorption of mineral and organic compounds and

415 the interaction of the concrete with seawater, which contains high concentrations of several

416 dissolved species (Vavrenyuk, Efimenko and Vavrenyuk, 2020). De Weerdt, Justnes and Geiker

417 (2014) have shown that, within about 20 days, cement-based binders immersed in seawater undergo

418 phase changes, whatever the type of binder tested (De Weerdt, Justnes and Geiker, 2014; Jakobsen,

419 De Weerdt and Geiker, 2016).

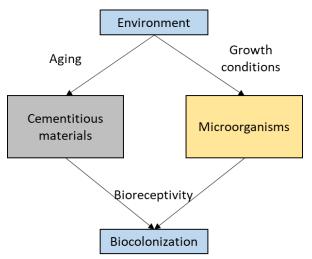


Figure 2. Interactions between the environment, microorganisms and cementitious materials
 (Lors, Feugeas and Tribollet, 2019)

423 There are a number of environmental factors that act on the colonization of cementitious materials in the marine environment, including luminosity, pH, temperature, nutrients and hydrodynamic 424 conditions (Azevedo, Carloni and Carvalheira, 2006; Choi et al., 2006; Färber et al., 2015; Firth et 425 al., 2014; Qvarfordt et al., 2006; Sherman, Giliam and Spieler, 2001; Witt et al., 2012). Of these, 426 it seems that the availability of nutrients and the temperature are the most likely factors to differ 427 428 between marine environments. Thus, these are considered the major factors influencing the extrinsic bioreceptivity of cementitious materials in these contexts (Färber et al., 2015; Gong, Song 429 and Warren 2005; Salta et al., 2013). 430

431 Nutrients availability

420

The medium is the means of propagation of nutrients allowing microorganisms to develop. The environment can be the source of nutrients, as is the case in sanitation networks, where the environment consists of wastewater rich in nutrients such as organic matter and sulfur compounds. Or the medium can simply be a means of transporting nutrients through the dissolution of organic matter. The concentration of nutrients present in the environment may favor the development of certain bacterial species. In the case of cementitious materials, these do not provide nutrients to the microorganisms, so the medium must be the source of nutrients to allow bacterial growth.

439 Seawater contains micro- and macronutrients essential for the growth of the cells of living 440 organisms. Macronutrients, which contain carbon (C), phosphorus (P), nitrogen (N) and silicon 441 (Si), provide the nutritive requirements indispensable to the metabolic functions of living 442 organisms. Micronutrients, containing iron (Fe), Zinc (Zn), Copper (Cu) or Manganese (Mn), while 443 consumed in much lower quantities, act as enzyme cofactors for metabolic reactions (de Fommervault, 2016). The existence of these nutrients in seawater make it rich not only in macroorganisms, but also microorganisms such as bacteria, cyanobacteria and microalgae (*Poli et al.*,
2017). The concentration and diversity of marine organisms also depend on other environmental
conditions, such as pressure, temperature and salinity. Biodiversity is lower where these conditions
are more drastic and hence more selective (Das, Lyla and Khan *et al.*, 2006; Sheehan, 2001;
Zeppilli *et al.*, 2018).

In laboratory tests, the use of artificial seawater as a culture medium has been shown to be sufficient
for marine bacteria to form a mature biofilm on a glass surface (Guillonneau *et al.*, 2018; Hayek *et al.*, 2019; Mireille Ayé *et al.*, 2015). This medium lacks organic substances, indicating that marine

453 bacteria can form a biofilm without the need organic matter.

454 **Temperature**

455 Temperature is one of the most important factors affecting the growth of microorganisms (Mesquita et al., 2020). The optimum temperature for most microorganisms is between 15°C and 456 35°C (Lors, Feugeas and Tribollet, 2019; Tran, 2011). A temperature that is too low reduces the 457 kinetics of the enzymatic reactions that condition the metabolism of microorganisms and thereby 458 their growth (González-Figueredo et al., 2018; Huston, Krieger-Brockett and Deming, 2000; 459 Robinson, 2015). A temperature that is too high damages microbial cells by denaturing the enzymes 460 and proteins of transport systems (Shehadul Islam, Aryasomayajula and Selvaganapathy, 2017). 461 However, certain microorganisms can resist temperatures close to 0°C and greater than or equal to 462 70°C (Brock, 2012). Moreover, each microorganism has an optimum growth temperature 463 dependent on its original environment (Lors, Feugeas and Tribollet, 2019). For example, marine 464 bacteria isolated from sediment, water or biofilm samples (adhered to a surface) are mesophilic, 465 growing best in moderate temperatures. Their optimum growth temperatures are between 25°C and 466 30°C, but they can grow at 20°C and withstand a temperature of 4°C (Brian-Jaisson et al., 2014; 467 Guillonneau et al., 2018; Hau and Gralnick, 2007; Hayek et al., 2019; Makemson et al., 1997; 468 Mireille Ayé et al., 2015). 469

Likewise, the temperature of the medium affects the bioreceptivity of cementitious materials through the mechanisms of adhesion between microorganisms and material. The surface tension of a liquid medium varies with temperature, which influences the three-way equilibrium between material, medium and microorganism and can thus affect microorganism adhesion. In the same effect category, Van der Waals forces also vary with temperature (Nedwell, 1999).

- 475
- 476

<u>RESULTS: BIOLOGICAL COLONIZATION AND THE</u> <u>**DURABILITY OF CONCRETE**</u>

The effect of the materials factors (cited below) on the intrinsic bioreceptivity of cement-based materials in the marine environment was studied in situ by Hayek *et al.*, (2021). In this paper, the effect of several parameters in the two main stages (bacterial biofilm and colonization by algae and other marine organisms) of cementitious material biocolonization in the marine environment was tested. The main parameters studied were i) the surface roughness, ii) the surface hydrophobicity and iii) the chemical composition. Figure 3 present the results obtained after 6 (Figure 3a) and 78

(Figure 3b) days of immersion in seawater. The results obtained in this study are in keeping with 483 the literature cited above, in which a similar effect of materials factors on the intrinsic 484 bioreceptivity has been reported. Based on the persistence of their significant effect (after 78 days 485 of immersion in seawater), the intrinsic parameters that support greater biocolonization are 486 classified from more to less effective in the following order; surface roughness (190 %) > chemical 487 composition (slag cement instead Portland cement) (136 %) > chemical composition (presence of 488 formwork oil) (106 %). Therefore, at the material scale, surface roughness is the most effective 489 factor in designing bioreceptive concrete that enhances marine biodiversity (Browne and Chapman 490 2011; Dafforn et al., 2015; Hayek et al., 2021). 491

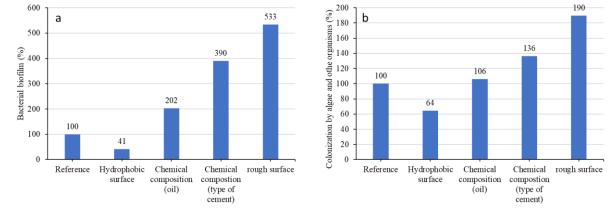




Figure 3. The effect of material factors on the intrinsic bioreceptivity of cement-based materials
in the marine environment. Data collected from the study of Hayek et al. (2021). (a) colonization
by bacteria after 6 days of immersion. (b) colonization by algae and other marine organisms after
78 days of immersion in seawater.

Hayek et al. (2020a) also showed that the carbonation of cementitious materials before immersion
in seawater plays a primary role in surface colonization by marine bacteria. Carbonated concrete
and mortar were shown to be more bioreceptive (200 % after 6 days of immersion in seawater)
than non-carbonated cement-based materials.

However, regarding the environmental factors, several authors have mentioned the importance of 501 nutrients availability. For example, in the case of artificial reef structures whose objective is the 502 colonization of the material, several 'pro-biofouling' coatings have been tried. The constituents of 503 these coatings are today known as chemo-attractors that allow microorganisms to adhere and grow 504 on the material (Lee et al., 2008; Souche et al., 2016). Lee et al. (2008) have found that the use of 505 506 chemoattractant coatings such as ferrous sulfate, zinc oxide, ammonium nitrate, sodium phosphates and ferrous lactate increase the colonization of cementitious materials in the marine environment. 507 A maximum biocolonization with green algae 25%, red algae 11.3% and brown algae 63.7% was 508

509 estimated from ferrous lactate coating.

510 Moreover, the colonization of cementitious material differs depending on the season (temperature 511 effect) (Choi *et al.*, 2006; Manso *et al.*, 2015). In a study carried out by Rica et al. (2016) focusing

512 on the effect of temperature on the colonizing biomass of a concrete surface in a marine

- environment, a decrease in temperature from 18°C to 13°C led to a regression (from 100% to 33%)
- in the biomass of the biofilm formed on concrete immersed in seawater.

515 Biological colonization can have positive or negative effects on a structure, either protecting or degrading the material (Coombes et al., 2011; Georges et al., 2021; Harshada, Desai and Gupta, 516 2017). Relatively little attention has been paid to the biodegradation of concrete in seawater, despite 517 the fact that the colonization of marine concrete structures is spontaneous. This can be explained 518 519 by the difficulty of separating biodeterioration from other processes that damage cementitious materials in seawater (e.g. aging, chemical erosion by chlorides, sulfate, etc.). In 2018, a laboratory 520 experiment demonstrated that the colonization of cementitious materials by microorganisms such 521 as the diatom Cylindrotheca closterium allowed greater penetration of chloride and sulfate ions, 522 which accelerated the degradation of the material. The author posited that the creation of cracks in 523 the material by diatoms could explain this phenomenon (Ferrero, 2018). 524

Yet as marine environments are extremely diverse, the results of experiments carried out in the 525 laboratory do not always reflect reality. A colonization effect determined in the laboratory is highly 526 dependent on the choice of microorganisms, the studied material (concrete, mortar, cement paste, 527 Portland cement, etc.) and the experimental conditions. According to Lv et al. (2015), the 528 colonization of concrete may actually improve its lifespan depending on the substrate and 529 microorganisms involved. These authors showed that the biofilm formed by bacteria of the genus 530 531 Pseudoalteromonas on the surface of the cement-based material improved its durability by reducing the quantity of aggressive ions such as Cl⁻ able to penetrate the material and cause 532 biodeterioration. This protective effect of biofilm has been identified not only in the laboratory 533 with bacteria, but also in situ with biofilm made up of other marine microorganisms and macro-534 organisms such as cyanobacteria, algae, lichens and barnacles (Bertron, 2014; Coombes et al., 535 2011; de la Rosa, Warke and Smith, 2013). In a number of studies, these living organisms have 536 been found to reduce the permeability of the cement-based material, resulting in greater durability 537 (Chlayon, Iwanami and Chijiwa, 2018; Coombes et al., 2017; Patil et al., 1994; Perkol-Finkel and 538 Sella, 2014; Soleimani et al., 2013). 539

In this way, the biological colonization of concrete could provide structures with additional 540 protection against chemical or even mechanical degradation, through the effects of the physical 541 barrier offered by living organisms colonizing the surface of cementitious material (Ariño et al., 542 1995; Bartoli et al., 2014; Carter and Viles, 2003; Chlayon, Iwanami and Chijiwa, 2020; 543 Mottershead and Lucas, 2000; Viles and Cutler, 2012). According to the literature, a beneficial 544 biofilm (bioprotection) can protect the material by reducing the quantity of water and nutrients 545 available, synthesizing anti-microbial agents (thus inhibiting the growth of other microorganisms 546 having a harmful effect on cementitious materials) or forming a protective layer of synthesized 547 exopolysaccharides (Chlayon et al., 2018; Soleimani et al., 2013). 548

A microbial biofilm can also facilitate the attachment of other living organisms to the surface of the structure. In the case of cementitious materials, the attachment of living organisms such as oysters, tubeworms, barnacles and corals can lead to the formation of calcite precipitated in the gaps of the cement matrix, making the concrete denser and stronger (Chlayon *et al.*, 2018; Joshi,

553 Goyal and Reddy, 2018; Risinger, 2012).

554

555DISCUSSION: ECOLOGICAL CONSIDERATIONS AND556PERSPECTIVES TO DESIGN MORE BIORECEPTIVE557CONCRETE (GREEN CONCRETE)

It has been shown that the marine diversity decreases with the loss of coastal wetlands and 558 nearshore communities when marine infrastructure such as seawalls are constructed. The loss of 559 horizontal extent of coastal communities occurs with seawalls, contributing to poor ecological 560 performance, exacerbated by increased land-based sources of pollution. One of the major reasons 561 for the poor ecological performance of the marine infrastructure is the structural design which is 562 less complex than the natural communities replaced (Hosseinzadeh et al., 2022). However, if the 563 marine infrastructure is designed considering ecological features, natural marine habitats could be 564 replicated which supports some species of marine life (Dyson and Yocom, 2015). 565

566 While ideally coastal development should be minimized or avoided, as artificialization is the primary cause of biodiversity loss in the world (IPBES, 2019), in the short term the global demand 567 for this is set to continue. To reduce the environmental impact and the disturbance of coastal 568 ecosystems, we recommend using a new design method for coastal and marine structures. One way 569 to do this is to consider the use of bioreceptive concrete at the site of impact, conceived with the 570 optimal surface characteristics for the local ecosystem. This new design approach would aim to 571 enhance biological colonization of coastal engineering structures and should be considered at the 572 project outset (Pioch et al., 2018). The geometry or the physical shape of marine infrastructure is 573 one of the parameters that show a direct ecological effect; fishes and benthic invertebrates populate 574 and interact in greater numbers on materials with rugose and irregular surfaces rather than a 575 smooth, flat wall (Hosseinzadeh et al., 2022; Koraim, 2013; Perkol-Finkel and Sella, 2014; Yip, 576 Zhang and Chwang, 2002). 577

578 Moreover, based in the present paper and the literature cited above, the following recommendations 579 aim to meet the objective of the new design approach:

- 580 1. Map the ecosystem at the site where the structure will be built and identify sensitive species.
- 581 2. Test the compatibility of the proposed concrete composition with the ecosystem where the
 582 project is planned, using the test method proposed by Hayek *et al.* (2020a).
- 583 3. Design surface roughness to ensure adhesion.
- 4. Pretreat the surface (e.g. carbonation), if possible, to accelerate growth.
- 585 5. Adjust the timing of the project (e.g. season) so that the biophysical conditions (dependent 586 on the area) are favorable for the ecosystem to start colonization quickly.
- 587 6. Develop a monitoring plan to ensure the reestablishment of the ecosystem after588 construction, as well as a solution to fight against invasive species if needed.
- 589 7. Do not apply a hydrophobic coating such as the curing product on the concrete surface.
- 590 8. Use slag cement (CEM III) instead of Portland cement (CEM I) to prepare the concrete.
- 591

593

CONCLUSIONS

This review of literature presents some of the latest research results that contributes to enhancing 594 the colonization of marine infrastructure built with concrete. Biological colonization can reduce 595 596 negative impacts on natural and local ecosystems and lower ubiquitous exotic species invasion. To encourage surface colonization by marine microorganisms, the carbonation of cementitious 597 materials before their immersion in seawater play the primary role. The type of cement influences 598 the kinetics and the level of surface colonization. Moreover, latest findings clearly indicate that 599 slag cement (CEM III) enhances marine biodiversity as compared to Portland cement (CEM I). At 600 the material scale, surface roughness is the most effective factor in designing bioreceptive concrete 601 that enhances marine biodiversity. Based on the persistence of their significant effect (after 78 days 602 603 of immersion in seawater), the intrinsic parameters that support greater biocolonization are 604 classified from more to less effective in the following order; surface roughness, CEM III as binder, 605 chemical composition (presence of formwork oil).

But, adopting new methods of construction often requires overcoming a certain inertia: insurance 606 and construction companies, as well as resource managers, tend to try to avoid technical issue or 607 608 legal repercussions by repeating historically proven methods, ignoring the ecological aspect or hiding negatives impacts. Nonetheless, there is no question that coastal areas need to be considered 609 610 as tremendous reservoirs of biodiversity (IPBES, 2019). Ecological engineering inspiration to eco-611 design concrete infrastructures, is one way to mitigate negative impact from human construction, 612 and should help to restore marine biodiversity which could be saved by 2050, if appropriate policies 613 and projects in the field start quickly (Duarte *et al.*, 2020). This review highlights some of the most significant results demonstrating that concrete can be an appropriate support for biological 614 615 colonization, making it a candidate for green design strategies. Of course, such an approach should not be used as a 'greenwashing' excuse to allow unregulated construction anywhere, thus becoming 616 a Trojan horse to ongoing massive artificialization (Firth et al., 2020). For real and fair progress to 617 be made, it will be essential to develop robust assessment methods to measure the ecological gains 618 of bioreceptive designed infrastructure and prove their potential benefits. 619

However, to increase the understanding of biofilm formation on concrete surfaces in the marine environment and to analyses the ecological benefits of this biocolonization, further investigation is needed using the molecular biology tools such as the amplicon sequencing of the 16S rRNA gene in order to investigate succession of the biofilm communities and their interaction with the concrete material and the flowing water.

625

626

- **ACKNOWLEDGMENTS**
- The authors are grateful to IMT Mines d'Alès and University of Paul Valéry Montpellier 3 for
 their participation and support.
- 629

630	
631	LITERATURE CITED
632	Al-Handal, A.Y., Wulff, A., 2008. Marine epiphytic diatoms from the shallow sublittoral zone in
633	Potter Cove, King George Island, Antarctica. Bot. Mar. 51, 411–435.
634	https://doi.org/10.1515/BOT.2008.053
635	Ammar, Y., Swailes, D., Bridgens, B., Chen, J., 2015. Influence of surface roughness on the initial
636	formation of biofilm. Surf. Coat. Technol., The 42nd International Conference on
637	Metallurgical Coatings and Thin Films 284, 410–416.
638	https://doi.org/10.1016/j.surfcoat.2015.07.062
639	An, Y.H., Friedman, R.J., 1998. Concise review of mechanisms of bacterial adhesion to biomaterial
640	surfaces. J. Biomed. Mater. Res. 43, 338-348. https://doi.org/10.1002/(SICI)1097-
641	4636(199823)43:3<338::AID-JBM16>3.0.CO;2-B
642	Anderson, M.J., Underwood, A.J., 1994. Effects of substratum on the recruitment and development
643	of an intertidal estuarine fouling assemblage. J. Exp. Mar. Biol. Ecol. 184, 217–236.
644	https://doi.org/10.1016/0022-0981(94)90006-X
645	Ariño, X., Ortega-Calvo, J.J., Gomez-Bolea, A., Saiz-Jimenez, C., 1995. Lichen colonization of
646	the Roman pavement at Baelo Claudia (Cadiz, Spain): biodeterioration vs. bioprotection.
647	Sci. Total Environ., The Deterioration of Monuments 167, 353–363.
648	https://doi.org/10.1016/0048-9697(95)04595-R
649	Azevedo, Flávia Beatriz Beserra, Carloni, G.G., Carvalheira, L.V., 2006. Colonization of benthic
650	organisms on different artificial substratum in Ilha Grande bay, Rio de Janeiro, Brazil. Braz.
651	Arch. Biol. Technol. 49, 263–275. https://doi.org/10.1590/S1516-89132006000300012
652	Azevedo, F. B B, Carloni, G.G., Carvalheira, L. V., 2006. Colonization of benthic organisms on
653	different artificial substratum in Ilha Grande Bay, Rio de Janeiro, Brazil. Braz. Arch. Biol.
654 655	Technol. 49, 263–275. https://doi.org/10.1590/S1516-89132006000300012 Baine, M., 2001. Artificial reefs: a review of their design, application, management and
656	performance. Ocean Coast. Manag. 44, 241–259. https://doi.org/10.1016/S0964-
657	5691(01)00048-5
658	Barraud, N., Schleheck, D., Klebensberger, J., Webb, J.S., Hassett, D.J., Rice, S.A., Kjelleberg, S.,
659	2009. Nitric Oxide Signaling in Pseudomonas aeruginosa Biofilms Mediates
660	Phosphodiesterase Activity, Decreased Cyclic Di-GMP Levels, and Enhanced Dispersal. J.
661	Bacteriol. 191, 7333–7342. https://doi.org/10.1128/JB.00975-09
662	Bartoli, F., Municchia, A.C., Futagami, Y., Kashiwadani, H., Moon, K.H., Caneva, G., 2014.
663	Biological colonization patterns on the ruins of Angkor temples (Cambodia) in the
664	biodeterioration vs bioprotection debate. Int. Biodeterior. Biodegrad. 96, 157–165.
665	https://doi.org/10.1016/j.ibiod.2014.09.015
666	Bellon-Fontaine, MN., Benezech, T., Boutroux, K., Hermon, C., 2016. Conception hygiénique
667	de matériel et nettoyage-désinfection. p. 12.
668	Bengtson, S., Sallstedt, T., Belivanova, V., Whitehouse, M., 2017. Three-dimensional preservation
669	of cellular and subcellular structures suggests 1.6 billion-year-old crown-group red algae.
670	PLOS Biol. 15, e2000735. https://doi.org/10.1371/journal.pbio.2000735
671	Berne, C., Ellison, C.K., Ducret, A., Brun, Y.V., 2018. Bacterial adhesion at the single-cell level.
672	Nat. Rev. Microbiol. 16, 616–627. https://doi.org/10.1038/s41579-018-0057-5

- Bertron, A., 2014. Understanding interactions between cementitious materials and microorganisms: a key to sustainable and safe concrete structures in various contexts.
 Mater. Struct. 47, 1787–1806. https://doi.org/10.1617/s11527-014-0433-1
- Bixlar Gregory, D., Bhushan, B., 2012. Biofouling: lessons from nature. Math. Phy. and Eng. Sci.
 370, 2381–2417. https://doi.org/10.1098/rsta.2011.0502
- Blainey, B.L., Marshall, K.C., 1991. The use of block copolymers to inhibit bacterial adhesion and
 biofilm formation on hydrophobic surfaces in marine habitats. Biofouling 4, 309–318.
 https://doi.org/10.1080/08927019109378221
- Boyd, A., Chakrabarty, A.M., 1994. Role of alginate lyase in cell detachment of Pseudomonas
 aeruginosa. Appl Env. Microbiol 60, 2355–2359.
- Branda, S.S., Vik, S., Friedman, L., Kolter, R., 2005. Biofilms: the matrix revisited. Trends
 Microbiol. 13, 20–26. https://doi.org/10.1016/j.tim.2004.11.006
- Briand, J.-F., Pochon, X., Wood, S.A., Bressy, C., Garnier, C., Réhel, K., Urvois, F., Culioli, G.,
 Zaiko, A., 2018. Metabarcoding and metabolomics offer complementarity in deciphering
 marine eukaryotic biofouling community shifts. Biofouling 34, 657–672.
 https://doi.org/10.1080/08927014.2018.1480757
- Brian-Jaisson, F., Ortalo-Magné, A., Guentas-Dombrowsky, L., Armougom, F., Blache, Y.,
 Molmeret, M., 2014. Identification of Bacterial Strains Isolated from the Mediterranean
 Sea Exhibiting Different Abilities of Biofilm Formation. Microb. Ecol. 68, 94–110.
 https://doi.org/10.1007/s00248-013-0342-9
- Brock, T.D., 2012. Thermophilic Microorganisms and Life at High Temperatures. Springer
 Science & Business Media.
- Browne, M.A., Chapman, M.G., 2011. Ecologically Informed Engineering Reduces Loss of
 Intertidal Biodiversity on Artificial Shorelines. Environ. Sci. Technol. 45, 8204–8207.
 https://doi.org/10.1021/es201924b
- Bugnot, A.B., Mayer-Pinto, M., Airoldi, L., Heery, E.C., Johnston, E.L., Critchley, L.P., Strain,
 E.M.A., Morris, R.L., Loke, L.H.L., Bishop, M.J., Sheehan, E.V., Coleman, R.A., Dafforn,
 K.A., 2020. Current and projected global extent of marine built structures. Nat. Sustain. 1–
 9. https://doi.org/10.1038/s41893-020-00595-1
- Bulleri, F., Airoldi, L., 2005. Artificial marine structures facilitate the spread of a non-indigenous green alga, Codium fragile ssp. tomentosoides, in the north Adriatic Sea. J. Appl. Ecol. 42, 1063–1072. https://doi.org/10.1111/j.1365-2664.2005.01096.x
- Bulleri, F., Chapman, M.G., 2010. The introduction of coastal infrastructure as a driver of change
 in marine environments. J. Appl. Ecol. 47, 26–35. https://doi.org/10.1111/j.13652664.2009.01751.x
- Bulleri, F., Chapman, M.G., 2004. Intertidal assemblages on artificial and natural habitats in 708 Biol. 381-391. 709 marinas on the north-west coast of Italy. Mar. 145. 710 https://doi.org/10.1007/s00227-004-1316-8
- Bunt, C.R., Jones, D.S., Tucker, I.G., 1993. The effects of pH, ionic strength and organic phase on
 the bacterial adhesion to hydrocarbons (BATH) test. Int. J. Pharm. 99, 93–98.
 https://doi.org/10.1016/0378-5173(93)90350-O
- Busscher, H.J., Weerkamp, A.H., 1987. Specific and non-specific interactions in bacterial adhesion
 to solid substrata. FEMS Microbiol. Rev. 3, 165–173. https://doi.org/10.1111/j.15746968.1987.tb02457.x

- Carpentier, B., Cerf, O., 1993. Biofilms and their consequences, with particular reference to
 hygiene in the food industry. J. Appl. Bacteriol. 75, 499–511.
 https://doi.org/10.1111/j.1365-2672.1993.tb01587.x
- Carter, N.E.A., Viles, H.A., 2003. Experimental investigations into the interactions between
 moisture, rock surface temperatures and an epilithic lichen cover in the bioprotection of
 limestone. Build. Environ., Building Stone Decay: Observations, Experiments and
 Modeling 38, 1225–1234. https://doi.org/10.1016/S0360-1323(03)00079-9
- Chao, L., Rakshe, S., Leff, M., Spormann, A.M., 2013. PdeB, a cyclic Di-GMP-specific
 phosphodiesterase that regulates Shewanella oneidensis MR-1 motility and biofilm
 formation. J. Bacteriol. 195, 3827–3833. https://doi.org/10.1128/JB.00498-13
- Chapman, M.G., Underwood, A.J., 2011. Evaluation of ecological engineering of "armoured"
 shorelines to improve their value as habitat. J. Exp. Mar. Biol. Ecol., Global change in
 marine ecosystems 400, 302–313. https://doi.org/10.1016/j.jembe.2011.02.025
- Chatterjee, A., Goyns, A., 2013. Cementitious Materials Performance in Aggressive Aqueous
 Environments Engineering Perspectives, in: Alexander, M., Bertron, A., De Belie, N.
 (Eds.), Performance of Cement-Based Materials in Aggressive Aqueous Environments:
 State-of-the-Art Report, RILEM TC 211 PAE, RILEM State-of-the-Art Reports. Springer
 Netherlands, Dordrecht, pp. 391–449. https://doi.org/10.1007/978-94-007-5413-3_16
- Chlayon, T., Iwanami, M., Chijiwa, N., 2020. Impacts from concrete microstructure and surface
 on the settlement of sessile organisms affecting chloride attack. Constr. Build. Mater. 239,
 117863. https://doi.org/10.1016/j.conbuildmat.2019.117863
- Chlayon, T., Iwanami, M., Chijiwa, N., 2018. Combined protective action of barnacles and biofilm
 on concrete surface in intertidal areas. Constr. Build. Mater. 179, 477–487.
 https://doi.org/10.1016/j.conbuildmat.2018.05.223
- Choi, C., Ohno, M., Sohn, C., 2006. Algal succession on different substrata covering the artificial
 iron reef at Ikata in Shikoku, Japan. Algae 21, 305–310.
 https://doi.org/10.4490/ALGAE.2006.21.3.305
- Choi, C.G., Ohno, M., Sohn, C.H., 2006. Algal Succession on Different Substrata Covering the
 Artificial Iron Reef at Ikata in Shikoku, Japan. ALGAE 21, 305–310.
 https://doi.org/2006.21.3.305
- Codina, M., 2007. Les bétons bas pH Formulation, caractérisation et étude à long terme (Theses).
 INSA de Toulouse.
- Cooke, S.J., Bergman, J.N., Nyboer, E.A., Reid, A.J., Gallagher, A.J., Hammerschlag, N., Van de
 Riet, K., Vermaire, J.C., 2020. Overcoming the concrete conquest of aquatic ecosystems.
 Biol. Conserv. 247, 108589. https://doi.org/10.1016/j.biocon.2020.108589
- Coombes, M.A., Naylor, L.A., Thompson, R.C., Roast, S.D., Gómez-Pujol, L., Fairhurst, R.J.,
 2011. Colonization and weathering of engineering materials by marine microorganisms: an
 SEM study. Earth Surf. Process. Landf. 36, 582–593. https://doi.org/10.1002/esp.2076
- Coombes, M.A., Naylor, L.A., Viles, H.A., Thompson, R.C., 2013. Bioprotection and disturbance:
 Seaweed, microclimatic stability and conditions for mechanical weathering in the intertidal
 zone. Geomorphology 202, 4–14. https://doi.org/10.1016/j.geomorph.2012.09.014
- Coombes, M.A., Viles, H.A., Naylor, L.A., La Marca, E.C., 2017. Cool barnacles: Do common biogenic structures enhance or retard rates of deterioration of intertidal rocks and concrete?
 Sci. Total Environ. 580, 1034–1045. https://doi.org/10.1016/j.scitotenv.2016.12.058
- Costa, A., Appleton, J., 2001. Concrete carbonation and chloride penetration in a marine
 environment. Concr. Sci. Eng. 3, 8.

- Crutzen, P.J., 2006. The "Anthropocene," in: Ehlers, E., Krafft, T. (Eds.), Earth System Science in
 the Anthropocene. Springer, Berlin, Heidelberg, pp. 13–18. https://doi.org/10.1007/3-54026590-2_3
- 766 Cwalina, B., 2008. Biodeterioration of concrete. Archit. Civ. Eng. Environ. 4, 133–140.
- Dafforn, K.A., Glasby, T.M., Airoldi, L., Rivero, N.K., Mayer-Pinto, M., Johnston, E.L., 2015.
 Marine urbanization: an ecological framework for designing multifunctional artificial structures. Front. Ecol. Environ. 13, 82–90.
- Dalod, E., Govin, A., Guyonnet, R., Grosseau, P., Lors, C., Damidot, D., 2014. t, in: Fentiman,
 K.L., C.H.;. Mangabhai, R.J.;. Scrivener (Ed.), International Conference on Calcium
 Aluminates. IHS BRE Press, Palais des Papes, Avignon, France, pp. 523–534.
- Dang, H., Lovell, C.R., 2015. Microbial Surface Colonization and Biofilm Development in Marine
 Environments. Microbiol. Mol. Biol. Rev. MMBR 80, 91–138.
 https://doi.org/10.1128/MMBR.00037-15
- Das, S., Lyla, P.S., Khan, S.A., 2006. Marine microbial diversity and ecology: importance and
 future perspectives. Curr. Sci. 90, 1325–1335.
- De Belie, N., 2010. Microorganisms versus stony materials: a love–hate relationship. Mater. Struct.
 43, 1191–1202.
- De Fommervault, O.P., 2016. Dynamique des nutriments en Méditerranée: des campagnes
 océanographiques aux flotteurs Bio-Argo.
- De Fraiture, C., Wichelns, D., Rockstrom, J., Kemp-Benedict, E., Eriyagama, N., Gordon, L.J.,
 Hanjra, M.A., Hoogeveen, J., Huber-Lee, A., Karlberg, L., 2007. Looking ahead to 2050:
 scenarios of alternative investment approaches.
- De la Rosa, J.P.M., Warke, P.A., Smith, B.J., 2013. Lichen-induced biomodification of calcareous
 surfaces: Bioprotection versus biodeterioration. Prog. Phys. Geogr. Earth Environ. 37, 325–
 351. https://doi.org/10.1177/0309133312467660
- De Weerdt, K., Justnes, H., Geiker, M.R., 2014. Changes in the phase assemblage of concrete
 exposed to sea water. Cem. Concr. Compos., Special issue: Durability of concrete 47, 53–
 63. https://doi.org/10.1016/j.cemconcomp.2013.09.015
- Dobretsov, S., Rittschof, D., 2020. Love at First Taste: Induction of Larval Settlement by Marine
 Microbes. Int. J. Mol. Sci. 21, 731. https://doi.org/10.3390/ijms21030731
- Dobretsov, S., Teplitski, M., Bayer, M., Gunasekera, S., Proksch, P., Paul, V.J., 2011. Inhibition
 of marine biofouling by bacterial quorum sensing inhibitors. Biofouling 27, 893–905.
 https://doi.org/10.1080/08927014.2011.609616
- Dobretsov, S., Teplitski, M., Paul, V., 2009. Mini-review: quorum sensing in the marine
 environment and its relationship to biofouling. Biofouling 25, 413–427.
 https://doi.org/10.1080/08927010902853516
- Donlan, R.M., Costerton, J.W., 2002. Biofilms: Survival Mechanisms of Clinically Relevant
 Microorganisms. Clin. Microbiol. Rev. 15, 167–193.
 https://doi.org/10.1128/CMR.15.2.167-193.2002
- B02 Doody, J.P., 2004. 'Coastal squeeze'— an historical perspective. J. Coast. Conserv. 10, 129–138.
 B03 https://doi.org/10.1652/1400-0350(2004)010[0129:CSAHP]2.0.CO;2
- Duarte, C.M., Agusti, S., Barbier, E., Britten, G.L., Castilla, J.C., Gattuso, J.-P., Fulweiler, R.W.,
 Hughes, T.P., Knowlton, N., Lovelock, C.E., Lotze, H.K., Predragovic, M., Poloczanska,
 E., Roberts, C., Worm, B., 2020. Rebuilding marine life. Nature 580, 39–51.
 https://doi.org/10.1038/s41586-020-2146-7

- Bubosc, A., Escadeillas, G., Blanc, P.J., 2001. Characterization of biological stains on external
 concrete walls and influence of concrete as underlying material. Cem. Concr. Res. 31,
 1613–1617. https://doi.org/10.1016/S0008-8846(01)00613-5
- Bunne, W.M., 2002. Bacterial Adhesion: Seen Any Good Biofilms Lately? Clin. Microbiol. Rev.
 15, 155–166. https://doi.org/10.1128/CMR.15.2.155-166.2002
- 813 Dürr, S., Thomason, J.C., 2009. Biofouling. John Wiley & Sons.
- Dyson, K., Yocom, K., 2015. Ecological design for urban waterfronts. Urban. Ecosyst. 18, 189–
 208. https://doi.org/10.1007/s11252-014-0385-9
- El-Naggar, M.Y., Wanger, G., Leung, K.M., Yuzvinsky, T.D., Southam, G., Yang, J., Lau, W.M., 816 817 Nealson, K.H., Gorby, Y.A., 2010. Electrical transport along bacterial nanowires from oneidensis MR-1. Proc. Shewanella Natl. Acad. Sci. 107. 18127-18131. 818 819 https://doi.org/10.1073/pnas.1004880107
- Escadeillas, G., Bertron, A., Blanc, P., Dubosc, A., 2007. Accelerated testing of biological stain
 growth on external concrete walls. Part 1: Development of the growth tests. Mater. Struct.
 40, 1061–1071. https://doi.org/10.1617/s11527-006-9205-x
- Escadeillas, G., Bertron, A., Ringot, E., Blanc, P.J., Dubosc, A., 2008. Accelerated testing of
 biological stain growth on external concrete walls. Part 2: Quantification of growths. Mater.
 Struct. 42, 937. https://doi.org/10.1617/s11527-008-9433-3
- Evans, K., Bax, N., Bernal, P., Corrales, M.B., Cryer, M., Forsterra, G., Gaymer, C.F.,
 Haussermann, V., Rice, J., 2016. D. South Pacific Ocean.
- Evans, K., Bax, N.J., Smith, D.C., 2018. Enhancing the robustness of a national assessment of the
 marine environment. Mar. Policy 98, 133–145.
 https://doi.org/10.1016/j.marpol.2018.08.011
- Färber, C., Wisshak, M., Pyko, I., Bellou, N., Freiwald, A., 2015. Effects of Water Depth, Seasonal
 Exposure, and Substrate Orientation on Microbial Bioerosion in the Ionian Sea (Eastern
 Mediterranean). PLOS ONE 10, e0126495. https://doi.org/10.1371/journal.pone.0126495
- Ferrario, F., Beck, M.W., Storlazzi, C.D., Micheli, F., Shepard, C.C., Airoldi, L., 2014. The
 effectiveness of coral reefs for coastal hazard risk reduction and adaptation. Nat. Commun.
 5, 3794. https://doi.org/10.1038/ncomms4794
- Ferrero, M.-A., 2018. Colonisation et biodétérioration des bétons en milieu marin : mise au point
 d'essais en laboratoire et influence de la composition chimique du matériau cimentaire
 (phdthesis). Normandie Université.
- Firth, L.B., Airoldi, L., Bulleri, F., Challinor, S., Chee, S.-Y., Evans, A.J., Hanley, M.E., Knights,
 A.M., O'Shaughnessy, K., Thompson, R.C., Hawkins, S.J., 2020. Greening of grey
 infrastructure should not be used as a Trojan horse to facilitate coastal development. J.
 Appl. Ecol. 57, 1762–1768. https://doi.org/10.1111/1365-2664.13683
- Firth, L.B., Thompson, R.C., Bohn, K., Abbiati, M., Airoldi, L., Bouma, T.J., Bozzeda, F.,
 Ceccherelli, V.U., Colangelo, M.A., Evans, A., Ferrario, F., Hanley, M.E., Hinz, H.,
 Hoggart, S.P.G., Jackson, J.E., Moore, P., Morgan, E.H., Perkol-Finkel, S., Skov, M.W.,
 Strain, E.M., van Belzen, J., Hawkins, S.J., 2014. Between a rock and a hard place:
 Environmental and engineering considerations when designing coastal defence structures.
 Coast. Eng., Coasts@Risks: THESEUS, a new wave in coastal protection 87, 122–135.
 https://doi.org/10.1016/j.coastaleng.2013.10.015
- Fitzhardinge, R.C., Bailey-Brock, J.H., 1989. Colonization of artificial reef materials by coral and
 other sessile organisms. Bull. Mar. Sci. 44, 567–579.

- Gaylarde, C.C., Gaylarde, P.M., 2005. A comparative study of the major microbial biomass of
 biofilms on exteriors of buildings in Europe and Latin America. Int. Biodeterior. Biodegrad.
 55, 131–139. https://doi.org/10.1016/j.ibiod.2004.10.001
- Georges, M., Bourguiba, A., Chateigner, D., Sebaibi, N., Boutouil, M., 2021. The study of long-term durability and bio-colonization of concrete in marine environment. Environ. Sustain.
 Indic. 10, 100120. https://doi.org/10.1016/j.indic.2021.100120
- Giannantonio, D.J., Kurth, J.C., Kurtis, K.E., Sobecky, P.A., 2009. Effects of concrete properties
 and nutrients on fungal colonization and fouling. Int. Biodeterior. Biodegrad. 63, 252–259.
 https://doi.org/10.1016/j.ibiod.2008.10.002
- Giraudel, C., Garcia, N., Ledoux, S., 2014. Single-layer breakwater armouring : feedback on the accropodeTM technology from site experience., in: Coastal Engineering 2014. pp. 1–15.
- Gong, J., Song, W., Warren, A., 2005. Periphytic ciliate colonization: annual cycle and responses
 to environmental conditions. Aquat. Microb. Ecol. 39, 159–170.
 https://doi.org/10.3354/ame039159
- González-Figueredo, C., Flores-Estrella, R.A., Rojas-Rejón, O.A., 2018. Fermentation:
 Metabolism, Kinetic Models, and Bioprocessing. Curr. Top. Biochem. Eng.
 https://doi.org/10.5772/intechopen.82195
- Govin, A., Tran, T.H., Guyonnet, R., Grosseau, P., Lors, C., Damidot, D., Devès, O., Ruot, B.,
 2013. Ability in biofouling by *Klebsormidium flaccidum* of mortars: Influence of the
 intrinsic characteristics. First Int. Conf. Concr. Sustain. ICCS S3-3–6 pp. 914–919.
- Gowell, M.R., Coombes, M.A., Viles, H.A., 2015. Rock-protecting seaweed? Experimental
 evidence of bioprotection in the intertidal zone. Earth Surf. Process. Landf. 40, 1364–1370.
 https://doi.org/10.1002/esp.3736
- Brosseau, P., Dalod, E., Govin, A., Lors, C., Guyonnet, R., Damidot, D., 2015. Effect of the
 Chemical Composition of Building Materials on Algal Biofouling.
- Guillitte, O., 1995. Bioreceptivity: a new concept for building ecology studies. Sci. Total Environ.,
 The Deterioration of Monuments 167, 215–220. https://doi.org/10.1016/00489697(95)04582-L
- Guillitte, O., Dreesen, R., 1995. Laboratory chamber studies and petrographical analysis as
 bioreceptivity assessment tools of building materials. Sci. Total Environ., The Deterioration
 of Monuments 167, 365–374. https://doi.org/10.1016/0048-9697(95)04596-S
- Guillonneau, R., Baraquet, C., Bazire, A., Molmeret, M., 2018. Multispecies Biofilm Development
 of Marine Bacteria Implies Complex Relationships Through Competition and Synergy and
 Modification of Matrix Components. Front. Microbiol. 9, 1960.
 https://doi.org/10.3389/fmicb.2018.01960
- Ha, D.-G., O'Toole, G.A., 2015. c-di-GMP and its Effects on Biofilm Formation and Dispersion:
 a Pseudomonas Aeruginosa Review. Microbiol. Spectr. 3, MB-0003-2014.
 https://doi.org/10.1128/microbiolspec.MB-0003-2014
- Harper, K.T., Pendleton, R.L., 1993. CYANOBACTERIA AND CYANOLICHENS: CAN THEY
 ENHANCE AVAILABILITY OF ESSENTIAL MINERALS FOR HIGHER PLANTS?
 Gt. Basin Nat. 53, 59–72.
- Harshada, M.K., Desai, P.M.M., Gupta, D.A.K., 2017. Deterioration of Concrete in Marine
 Structure.
- Hau, H.H., Gralnick, J.A., 2007. Ecology and Biotechnology of the Genus Shewanella. Annu. Rev.
 Microbiol. 61, 237–258. https://doi.org/10.1146/annurev.micro.61.080706.093257

- Hayek, M., Baraquet, C., Lami, R., Blache, Y., Molmeret, M., 2019. The Marine Bacterium
 Shewanella woodyi Produces C8-HSL to Regulate Bioluminescence. Microb. Ecol.
 https://doi.org/10.1007/s00248-019-01454-z
- Hayek, M., Salgues, M., Habouzit, F., Bayle, S., Souche, J.-C., De Weerdt, K., Pioch, S., 2020a.
 In vitro and in situ tests to evaluate the bacterial colonization of cementitious materials in
 the marine environment. Cem. Concr. Compos. 113, 103748.
 https://doi.org/10.1016/j.cemconcomp.2020.103748
- Hayek, M., Salgues, M., Habouzit, F., Bayle, S., Souche, J.-C., Weerdt, K.D., Pioch, S., 2020b.
 L'influence de la carbonatation sur la biocolonisation de matériaux cimentaires dans le
 milieu marin. Matér. Tech. 108, 202. https://doi.org/10.1051/mattech/2020020
- Hayek, M., Salgues, M., Souche, J.-C., Cunge, E., Giraudel, C., Paireau, O., 2021. Influence of the
 Intrinsic Characteristics of Cementitious Materials on Biofouling in the Marine
 Environment. Sustainability 13, 2625. https://doi.org/10.3390/su13052625
- Herrmann, M., Vaudaux, P.E., Pittet, D., Auckenthaler, R., Lew, P.D., Schumacher-Perdreau, F.,
 Peters, G., Waldvogel, F.A., 1988. Fibronectin, fibrinogen, and laminin act as mediators of
 adherence of clinical staphylococcal isolates to foreign material. J. Infect. Dis. 158, 693–
 701.
- Hinsa, S.M., O'Toole, G.A., 2006. Biofilm formation by Pseudomonas fluorescens WCS365: a
 role for LapD. Microbiol. Read. Engl. 152, 1375–1383.
 https://doi.org/10.1099/mic.0.28696-0
- Hosseinzadeh, N., Ghiasian, M., Andiroglu, E., Lamere, J., Rhode-Barbarigos, L., Sobczak, J.,
 Sullivan-Sealey, K., Suraneni, P., 2022. Concrete seawalls: A review of load
 considerations, ecological performance, durability, and recent innovations. Ecol. Eng. 178,
 106573. https://doi.org/10.1016/j.ecoleng.2022.106573
- Huggett, M.J., Nedved, B.T., Hadfield, M.G., 2009. Effects of initial surface wettability on biofilm
 formation and subsequent settlement of Hydroides elegans. Biofouling 25, 387–399.
 https://doi.org/10.1080/08927010902823238
- Huston, A.L., Krieger-Brockett, B.B., Deming, J.W., 2000. Remarkably low temperature optima for extracellular enzyme activity from Arctic bacteria and sea ice. Environ. Microbiol. 2, 383–388. https://doi.org/10.1046/j.1462-2920.2000.00118.x
- Jackson, J.B.C., Kirby, M.X., Berger, W.H., Bjorndal, K.A., Botsford, L.W., Bourque, B.J.,
 Bradbury, R.H., Cooke, R., Erlandson, J., Estes, J.A., Hughes, T.P., Kidwell, S., Lange,
 C.B., Lenihan, H.S., Pandolfi, J.M., Peterson, C.H., Steneck, R.S., Tegner, M.J., Warner,
 R.R., 2001. Historical Overfishing and the Recent Collapse of Coastal Ecosystems. Science
 293, 629–637. https://doi.org/10.1126/science.1059199
- Jakobsen, U.H., De Weerdt, K., Geiker, M.R., 2016. Elemental zonation in marine concrete. Cem.
 Concr. Res. 85, 12–27. https://doi.org/10.1016/j.cemconres.2016.02.006
- Joshi, S., Goyal, S., Reddy, M.S., 2018. Influence of nutrient components of media on structural
 properties of concrete during biocementation. Constr. Build. Mater. 158, 601–613.
 https://doi.org/10.1016/j.conbuildmat.2017.10.055
- Kaplan, J.B., 2010. Biofilm dispersal: mechanisms, clinical implications, and potential therapeutic
 uses. J. Dent. Res. 89, 205–218. https://doi.org/10.1177/0022034509359403
- Karunasagar, I., Otta, S.K., Karunasagar, I., Indrani, K., 1996. Biofilm formation by Vibrio harveyi
 on surfaces.

- Katsikogianni, M., Missirlis, Y., 2004. Concise review of mechanisms of bacterial adhesion to
 biomaterials and of techniques used in estimating bacteria-material interactions. Eur. Cell.
 Mater. 8, 37–57. https://doi.org/10.22203/eCM.v008a05
- Koraim, A.S., 2013. Hydrodynamic efficiency of suspended horizontal rows of half pipes used as
 a new type breakwater. Ocean. Eng. 64, 1–22.
 https://doi.org/10.1016/j.oceaneng.2013.02.008
- Krumbein, W.E., 1979. Photolithotropic and chemoorganotrophic activity of bacteria and algae as
 related to beachrock formation and degradation (gulf of Aqaba, Sinai). Geomicrobiol. J. 1,
 139–203. https://doi.org/10.1080/01490457909377729
- La Marca, E.C., Coombes, M.A., Viles, H.A., Naylor, L.A., 2014. THE BIO-PROTECTIVE ROLE
 OF A BIOLOGICAL ENCRUSTATION. Biol. Mar. Mediterr. 21, 345.
- Lacroix, D., Pioch, S., 2011. The multi-use in wind farm projects: more conflicts or a win-win
 opportunity? Aquat Living Resour EDP Sci. 24, 129–135.
 https://doi.org/10.1051/alr/2011135
- Lawrence, J.R., Scharf, B., Packroff, G., Neu, T.R., 2002. Microscale evaluation of the effects of grazing by invertebrates with contrasting feeding modes on river biofilm architecture and composition. Microb. Ecol. 44, 199–207. https://doi.org/10.1007/s00248-001-1064-y
- 959 Leclerc, H., 1975. Microbiologie générale, in: Microbiologie Generale. Doin.
- Lee, H.S., Sidharthan, M., Shim, C.S., Do Kim, Y., Lim, C.Y., Ko, J.W., Han, M.D., Rang, M.J.,
 Bim, L.S., Cho, H.S., 2008. Screening and formulation of chemoattractant coatings for
 artificial reef structures. J. Environ. Biol. 29, 605–612.
- 263 Loeb, G.I., Neihof, R.A., 1975. Marine conditioning films.
- Lors, C., Feugeas, F., Tribollet, B., 2019. Interactions Materials Microorganisms: Concrete and
 Metals more Resistant to Biodeterioration. EDP sciences.
- Lueangthuwapranit, C., Sampantarak, U., Wongsai, S., 2011. Distribution and Abundance of
 Phytoplankton: Influence of Salinity and Turbidity Gradients in the Na Thap River,
 Songkhla Province, Thailand. J. Coast. Res. 27, 585–594.
 https://doi.org/10.2112/JCOASTRES-D-10-00123.1
- Ly, O., Yoris-Nobile, A.I., Sebaibi, N., Blanco-Fernandez, E., Boutouil, M., Castro-Fresno, D.,
 Hall, A.E., Herbert, R.J.H., Deboucha, W., Reis, B., Franco, J.N., Teresa Borges, M.,
 Sousa-Pinto, I., van der Linden, P., Stafford, R., 2021. Optimisation of 3D printed concrete
 for artificial reefs: Biofouling and mechanical analysis. Constr. Build. Mater. 272, 121649.
 https://doi.org/10.1016/j.conbuildmat.2020.121649
- Makemson, J.C., Fulayfil, N.R., Landry, W., Van Ert, L.M., Wimpee, C.F., Widder, E.A., Case,
 J.F., 1997. Shewanella woodyi sp. nov., an Exclusively Respiratory Luminous Bacterium
 Isolated from the Alboran Sea. Int. J. Syst. Evol. Microbiol. 47, 1034–1039.
 https://doi.org/10.1099/00207713-47-4-1034
- Manso, S., Aguado, A., 2017. A review of sample preparation and its influence on pH
 determination in concrete samples. Mater. Constr. 67, 108.
 https://doi.org/10.3989/mc.2017.08515
- Manso, S., Aguado, A., 2016. The use of bio-receptive concrete as a new typology of living wall
 systems. Matér. Tech. 104, 502. https://doi.org/10.1051/mattech/2016028
- Manso, S., Calvo-Torras, M.Á., De Belie, N., Segura, I., Aguado, A., 2015. Evaluation of natural
 colonisation of cementitious materials: Effect of bioreceptivity and environmental
 conditions. Sci. Total Environ. 512–513, 444–453.
 https://doi.org/10.1016/j.scitotenv.2015.01.086

- Manso, S., De Muynck, W., Segura, I., Aguado, A., Steppe, K., Boon, N., De Belie, N., 2014.
 Bioreceptivity evaluation of cementitious materials designed to stimulate biological growth. Sci. Total Environ. 481, 232–241. https://doi.org/10.1016/j.scitotenv.2014.02.059
- Martins, G.M., Thompson, R.C., Neto, A.I., Hawkins, S.J., Jenkins, S.R., 2010. Enhancing stocks
 of the exploited limpet Patella candei d'Orbigny via modifications in coastal engineering.
 Biol. Conserv. 143, 203–211. https://doi.org/10.1016/j.biocon.2009.10.004
- McManus, R.S., Archibald, N., Comber, S., Knights, A.M., Thompson, R.C., Firth, L.B., 2018.
 Partial replacement of cement for waste aggregates in concrete coastal and marine
 infrastructure: A foundation for ecological enhancement? Ecol. Eng. 120, 655–667.
 https://doi.org/10.1016/j.ecoleng.2017.06.062
- Meijer, J.R., Huijbregts, M.A.J., Schotten, K.C.G.J., Schipper, A.M., 2018. Global patterns of
 current and future road infrastructure. Environ. Res. Lett. 13, 064006.
 https://doi.org/10.1088/1748-9326/aabd42
- Meng, H., Zhang, X., Katayama, Y., Ge, Q., Gu, J.-D., 2020. Microbial diversity and composition of the Preah Vihear temple in Cambodia by high-throughput sequencing based on genomic DNA and RNA. Int. Biodeterior. Biodegrad. 149, 104936. https://doi.org/10.1016/j.ibiod.2020.104936
- Mesquita, M.C.B., Prestes, A.C.C., Gomes, A.M.A., Marinho, M.M., 2020. Direct Effects of
 Temperature on Growth of Different Tropical Phytoplankton Species. Microb. Ecol. 79, 1–
 1007 11. https://doi.org/10.1007/s00248-019-01384-w
- Mieszkin, S., Callow, M.E., Callow, J.A., 2013. Interactions between microbial biofilms and marine fouling algae: a mini review. Biofouling 29, 1097–1113. https://doi.org/10.1080/08927014.2013.828712
- Miller, A.Z., Dionísio, A., Laiz, L., Macedo, M.F., Saiz-Jimenez, C., 2009. The influence of inherent properties of building limestones on their bioreceptivity to phototrophic microorganisms. Ann. Microbiol. 59, 705–713. https://doi.org/10.1007/BF03179212
- Miller, A.Z., Sanmartín, P., Pereira-Pardo, L., Dionísio, A., Saiz-Jimenez, C., Macedo, M.F.,
 Prieto, B., 2012. Bioreceptivity of building stones: A review. Sci. Total Environ. 426, 1–
 1016 12. https://doi.org/10.1016/j.scitotenv.2012.03.026
- Mireille Ayé, A., Bonnin-Jusserand, M., Brian-Jaisson, F., Ortalo-Magné, A., Culioli, G., Koffi
 Nevry, R., Rabah, N., Blache, Y., Molmeret, M., 2015. Modulation of violacein production
 and phenotypes associated with biofilm by exogenous quorum sensing N-acylhomoserine
 lactones in the marine bacterium Pseudoalteromonas ulvae TC14. Microbiology, 161,
 2039–2051. https://doi.org/10.1099/mic.0.000147
- Mottershead, D., Lucas, G., 2000. THE ROLE OF LICHENS IN INHIBITING EROSION OF A
 SOLUBLE ROCK. The Lichenologist 32, 601–609. https://doi.org/10.1006/lich.2000.0300
- Naylor, L., Coombes, M.A., 2015. Bioprotection: working with nature to manage coastal hazards.
 Geogr. Rev. 28, 37–41.
- Nedwell, D.B., 1999. Effect of low temperature on microbial growth: lowered affinity for
 substrates limits growth at low temperature. FEMS Microbiol. Ecol. 30, 101–111.
 https://doi.org/10.1111/j.1574-6941.1999.tb00639.x
- Nuhoglu, Y., Oguz, E., Uslu, H., Ozbek, A., Ipekoglu, B., Ocak, I., Hasenekoglu, I., 2006. The accelerating effects of the microorganisms on biodeterioration of stone monuments under air pollution and continental-cold climatic conditions in Erzurum, Turkey. Sci. Total Environ. 364, 272–283. https://doi.org/10.1016/j.scitotenv.2005.06.034

- 1033 Ogunola, O.S., Onada, O.A., 2016. Anti-Biofouling Defence Mechanism of Basibionts (A
 1034 Chemical Warfare) A Critical Review. J. Environ. Anal. Toxicol. 6, 1–6.
 1035 https://doi.org/10.4172/2161-0525.1000380
- O'Toole, G., Kaplan, H.B., Kolter, R., 2000. Biofilm formation as microbial development. Annu.
 Rev. Microbiol. 54, 49–79. https://doi.org/10.1146/annurev.micro.54.1.49
- Ottelé, M., Perini, K., Fraaij, A.L.A., Haas, E.M., Raiteri, R., 2011. Comparative life cycle analysis
 for green façades and living wall systems. Energy Build. 43, 3419–3429.
 https://doi.org/10.1016/j.enbuild.2011.09.010
- Parker, C.D., 1945. The corrosion of concrete: 1. The isolation of a species of bacterium associated
 with the corrosion of concrete exposed to atmospheres containing hydrogen sulphide. Aust.
 J. Exp. Biol. Med. Sci. 23, 81–90.
- Pasmore, M., Costerton, J.W., 2003. Biofilms, bacterial signaling, and their ties to marine biology.
 J. Ind. Microbiol. Biotechnol. 30, 407–413. https://doi.org/10.1007/s10295-003-0069-6
- Patil, B.T., Gajendragad, M.R., Ranganna, G., Wagh, A.B., Sudhakaran, T., 1994. Impact of
 biofouling on corrosion resistance of reinforced concrete.
- Patranella, A., Kilfoyle, K., Pioch, S., Spieler, R.E., 2017. Artificial reefs as juvenile fish habitat
 in a marina. J. Coast. Res. 33, 1341–1351.
- Perini, K., Ottelé, M., 2014. Designing green façades and living wall systems for sustainable
 constructions. Int. J. Des. Nat. Ecodynamics 9, 31–46. https://doi.org/10.2495/DNE-V9 N1-31-46
- Perkol-Finkel, S., Hadary, T., Rella, A., Shirazi, R., Sella, I., 2018. Seascape architecture incorporating ecological considerations in design of coastal and marine infrastructure. Ecol.
 Eng. 120, 645–654. https://doi.org/10.1016/j.ecoleng.2017.06.051
- Perkol-Finkel, S., Sella, I., 2014. Ecologically Active Concrete for Coastal and Marine
 Infrastructure: Innovative Matrices and Designs, in: From Sea to Shore? Meeting the
 Challenges of the Sea. ICE Publishing, pp. 1139–1149.
 https://doi.org/10.1680/fsts.59757.124
- Phan, L.K., van Thiel de Vries, J.S.M., Stive, M.J.F., 2015. Coastal Mangrove Squeeze in the
 Mekong Delta. J. Coast. Res. 31, 233–243. https://doi.org/10.2112/JCOASTRES-D-1400049.1
- Pioch, S., Kilfoyle, K., Levrel, H., Spieler, R., 2011. Green Marine Construction. J. Coast. Res. 61,
 257–268. https://doi.org/10.2112/SI61-001.1
- Pioch, S., Relini, G., Souche, J.C., Stive, M.J.F., De Monbrison, D., Nassif, S., Simard, F.,
 Allemand, D., Saussol, P., Spieler, R., Kilfoyle, K., 2018. Enhancing eco-engineering of
 coastal infrastructure with eco-design: Moving from mitigation to integration. Ecol. Eng.
 120, 574–584. https://doi.org/10.1016/j.ecoleng.2018.05.034
- Pioch, S., Souche, J.-C., 2021. L'écoconception des infrastructures maritimes: vers un aménagement intégré à l'environnement. ISTE Group.
- Poli, A., Finore, I., Romano, I., Gioiello, A., Lama, L., Nicolaus, B., 2017. Microbial Diversity in
 Extreme Marine Habitats and Their Biomolecules. Microorganisms 5.
 https://doi.org/10.3390/microorganisms5020025
- Poonguzhali, A., Shaikh, H., Dayal, R.K., Khatak, H.S., 2008. A Review on Degradation
 Mechanism and Life Estimation of Civil Structures. Corros. Rev. 26, 215–294.
 https://doi.org/10.1515/corrrev.2008.215

- Ovarfordt, S., Kautsky, H., Malm, T., 2006. Development of fouling communities on vertical 1077 the Baltic Sea. Estuar. Coast. Shelf Sci. 67. 618–628. 1078 structures in https://doi.org/10.1016/j.ecss.2006.01.004 1079
- 1080 Rajitha, K., Nancharaiah, Y.V., Venugopalan, V.P., 2020. Role of bacterial biofilms and their EPS
 1081 on settlement of barnacle (Amphibalanus reticulatus) larvae. Int. Biodeterior. Biodegrad.
 1082 150, 104958. https://doi.org/10.1016/j.ibiod.2020.104958
- 1083 Razavi, S.M.R., Oh, J., Haasch, R.T., Kim, K., Masoomi, M., Bagheri, R., Slauch, J.M., Miljkovic,
 1084 N., 2019. Environment-Friendly Antibiofouling Superhydrophobic Coatings. ACS Sustain.
 1085 Chem. Eng. 7, 14509–14520. https://doi.org/10.1021/acssuschemeng.9b02025
- 1086 Relini, G., Tixi, F., Relini, M., Torchia, G., 1998. The macrofouling on offshore platforms at
 1087 Ravenna. Int. Biodeterior. Biodegrad. 41, 41–55. https://doi.org/10.1016/S09641088 8305(98)80007-3
- Rica, H.C., Boutouil, M., Boudart, B., Claquin, P., Leroy, F., 2016. Colonisation et détérioration
 des bétons incorporant des coquilles pour récifs artificiels. Matér. Tech. 104, 503.
- Rindi, F., Guiry, M.D., López-Bautista, J.M., 2008. Distribution, morphology, and phylogeny of
 klebsormidium (Klebsormidiales, Charophyceae) in urban environments in europe 1. J.
 Phycol. 44, 1529–1540.
- 1094 Risinger, J., 2012. Biologically dominated engineered coastal breakwaters. LSU Dr. Diss.
- 1095 Robert, R., Chretiennot-Dinet, M.-J., Kaas, R., Martin-Jezequel, V., Moal, J., Le Coz, J.-R.,
 1096 Nicolas, J.-L., Bernard, E., Connan, J.-P., Le Dean, L., 2004. Amélioration des productions
 1097 phytoplanctoniques en écloserie de mollusques: caractérisation des microalgues fourrage.
- Roberts, D.J., Nica, D., Zuo, G., Davis, J.L., 2002. Quantifying microbially induced deterioration
 of concrete: initial studies. Int. Biodeterior. Biodegrad., Biodet.of Constr. Materials 49,
 227–234. https://doi.org/10.1016/S0964-8305(02)00049-5
- Robinson, P.K., 2015. Enzymes: principles and biotechnological applications. Essays Biochem.
 59, 1–41. https://doi.org/10.1042/bse0590001
- Romani, M., Carrion, C., Fernandez, F., Intertaglia, L., Pecqueur, D., Lebaron, P., Lami, R., 2019.
 High bacterial diversity in pioneer biofilms colonizing ceramic roof tiles. Int. Biodeterior.
 Biodegrad. 144, 104745. https://doi.org/10.1016/j.ibiod.2019.104745
- Salta, M., Wharton, J.A., Blache, Y., Stokes, K.R., Briand, J.-F., 2013. Marine biofilms on artificial
 surfaces: structure and dynamics. Environ. Microbiol. 15, 2879–2893.
 https://doi.org/10.1111/1462-2920.12186
- Scatigno, C., Moricca, C., Tortolini, C., Favero, G., 2016. The influence of environmental 1109 parameters in the biocolonization of the Mithraeum in the roman masonry of casa di Diana 1110 1111 (Ostia Antica, Italy). Environ. Sci. Pollut. Res. 23, 13403-13412. https://doi.org/10.1007/s11356-016-6548-x 1112
- Scheres, B., Schüttrumpf, H., 2019. Enhancing the Ecological Value of Sea Dikes. Water 11, 1617.
 https://doi.org/10.3390/w11081617
- Scheuerman, T.R., Camper, A.K., Hamilton, M.A., 1998. Effects of Substratum Topography on
 Bacterial Adhesion. J. Colloid Interface Sci. 208, 23–33.
 https://doi.org/10.1006/jcis.1998.5717
- Seto, K.C., Güneralp, B., Hutyra, L.R., 2012. Global forecasts of urban expansion to 2030 and direct impacts on biodiversity and carbon pools. Proc. Natl. Acad. Sci. 109, 16083–16088.
 https://doi.org/10.1073/pnas.1211658109
- Shearer, C.A., Descals, E., Kohlmeyer, B., Kohlmeyer, J., Marvanová, L., Padgett, D., Porter, D.,
 Raja, H.A., Schmit, J.P., Thorton, H.A., Voglymayr, H., 2007. Fungal biodiversity in

- 1123 aquatic habitats. Biodivers. Conserv. 16, 49–67. https://doi.org/10.1007/s10531-006-9120-1124 z
- 1125 Sheehan, P.M., 2001. History of marine biodiversity. Geol. J. 36, 231–249.
 1126 https://doi.org/10.1002/gj.890
- Shehadul Islam, M., Aryasomayajula, A., Selvaganapathy, P.R., 2017. A Review on Macroscale
 and Microscale Cell Lysis Methods. Micromachines 8. https://doi.org/10.3390/mi8030083
- Sherman, R.L., Gilliam, D.S., Spieler, R.E., 2001. Site-dependent differences in artificial reef
 function: implications for coral reef restoration. Bull. Mar. Sci. 69, 5.
- Shi, X., Zhu, X., 2009. Biofilm formation and food safety in food industries. Trends Food Sci.
 Technol., Natural and Safe Foods 20, 407–413. https://doi.org/10.1016/j.tifs.2009.01.054
- Shukla Jang, B., Maninder, S., Arora Maitri, V., Arvind, M., Yasuhiro, T., 2021. The Impact of
 Sea Level Rise Due to Global Warming on the Coastal Population Dynamics: A Modeling
 Study. Earth Systems and Environment. 5, 909–926. https://doi.org/10.1007/s41748-02100246-1
- 1137 Silva, M.R., Naik, T.R., 2013. Biodeterioration of Concrete Structures in Coastal Zone. p. 8.
- Soleimani, S., Ormeci, B., Isgor, O.B., 2013. Growth and characterization of Escherichia coli
 DH5α biofilm on concrete surfaces as a protective layer against microbiologically
 influenced concrete deterioration (MICD). Appl. Microbiol. Biotechnol. 97, 1093–1102.
- Somsueb, S., Ohno, M., Kimura, H., 2001. Development of seaweed communities on suspended
 substrata with three slope angles. J. Appl. Phycol. 13, 109–115.
 https://doi.org/10.1023/A:1011180527959
- Sosa, M., Pérez-López, T., Reyes, J., Corvo, F., Camacho-Chab, R., Quintana, P., Aguilar, D.,
 2011. Influence of the Marine Environment on Reinforced Concrete Degradation
 Depending on Exposure Conditions. Int J Electrochem Sci 6, 19.
- Souche, J.-C., Le Saout, G., Salgues, M., Pioch, S., 2016. Effets de bétons bio-actifs sur la colonisation marine en environnement méditerranéen. Matér. Tech. 104, 504.
- Souche, J.-C., Pioch, S., Salgues, M., Ienny, P., Le Saoût, G., 2018. interaction des ouvrages avec
 leur environnement. le milieu maritime, collection. ed. Paris.
- Souche, J.-C., Pioch, S., Salgues, M., Weerdt, K.D., Agostini, A., Hayek, M., 2019. De la
 conception à l'éco-conception des ouvrages maritimes : intégrer la nature au projet
 d'aménagement maritime. Rev. Paralia 12. https://doi.org/10.5150/revue-paralia.2019.n01
- Speranza, G., Gottardi, G., Pederzolli, C., Lunelli, L., Canteri, R., Pasquardini, L., Carli, E., Lui,
 A., Maniglio, D., Brugnara, M., Anderle, M., 2004. Role of chemical interactions in
 bacterial adhesion to polymer surfaces. Biomaterials 25, 2029–2037.
- Subashchandrabose, S.R., Ramakrishnan, B., Megharaj, M., Venkateswarlu, K., Naidu, R., 2011.
 Consortia of cyanobacteria/microalgae and bacteria: Biotechnological potential.
 Biotechnol. Adv. 29, 896–907. https://doi.org/10.1016/j.biotechadv.2011.07.009
- Tang, H., Wang, A., Liang, X., Cao, T., Salley, S.O., McAllister, J.P., Ng, K.Y.S., 2006. Effect of
 surface proteins on Staphylococcus Epidermidis adhesion and colonization on silicone.
 Colloids Surf. B Biointerfaces 51, 16–24. https://doi.org/10.1016/j.colsurfb.2006.04.011
- Temmerman, S., Meire, P., Bouma, T.J., Herman, P.M.J., Ysebaert, T., De Vriend, H.J., 2013.
 Ecosystem-based coastal defence in the face of global change. Nature 504, 79–83.
 https://doi.org/10.1038/nature12859
- Tomaselli, L., Lamenti, G., Bosco, M., Tiano, P., 2000. Biodiversity of photosynthetic micro organisms dwelling on stone monuments. Int. Biodeterior. Biodegrad. 46, 251–258.

- Tran, T.H., 2011. Influence of the intrinsic characteristics of a mortar on its ability to resist to the
 biofouling (Theses). Ecole Nationale Supérieure des Mines de Saint-Etienne.
- Tran, T.H., Govin, A., Guyonnet, R., Grosseau, P., Lors, C., Damidot, D., Devès, O., Ruot, B.,
 2014. Influence of the intrinsic characteristics of mortars on their biofouling by pigmented
 organisms: Comparison between laboratory and field-scale experiments. Int. Biodeterior.
 Biodegrad. 86, 334–342. https://doi.org/10.1016/j.ibiod.2013.10.005
- Tran, T.H., Govin, A., Guyonnet, R., Grosseau, P., Lors, C., Garcia-Diaz, E., Damidot, D., Devès,
 O., Ruot, B., 2012. Influence of the intrinsic characteristics of mortars on biofouling by
 Klebsormidium flaccidum, in: International Biodeterioration & Biodegradation. pp. 31–39.
 https://doi.org/doi.org/10.1016/j.ibiod.2011.10.017
- 1178 Valipour, M., Sayed M.B., Changhyun, J., 2021. Global Surface Temperature: A New Insight.
 1179 Climate, 9. https://doi.org/10.3390/cli9050081
- van Loosdrecht, M.C.M., Lyklema, J., Norde, W., Zehnder, A.J.B., 1989. Bacterial adhesion: A
 physicochemical approach. Microb. Ecol. 17, 1–15. https://doi.org/10.1007/BF02025589
- 1182 Vavrenyuk, S.V., Efimenko, Y.V., Vavrenyuk, V.G., 2020. On Standard Requirements to Cement
 1183 for Concrete Used in Marine Environment. IOP Conf. Ser. Mater. Sci. Eng. 753, 022040.
 1184 https://doi.org/10.1088/1757-899X/753/2/022040
- Vaz-Pinto, F., Torrontegi, O., Prestes, A.C.L., Alvaro, N. V., Neto, A.I., Martins, G.M., 2014. 1185 1186 Invasion success and development of benthic assemblages: Effect of timing, duration of submersion and substrate type. Mar. Environ. Res. 94. 72-79. 1187 https://doi.org/10.1016/j.marenvres.2013.12.007 1188
- 1189 Veeger, M., Ottelé, M., Prieto, A., 2021. Making bioreceptive concrete: Formulation and testing
 1190 of bioreceptive concrete mixtures. J. Build. Eng. 44, 102545.
 1191 https://doi.org/10.1016/j.jobe.2021.102545
- Viles, H.A., Cutler, N.A., 2012. Global environmental change and the biology of heritage
 structures. Glob. Change Biol. 18, 2406–2418. https://doi.org/10.1111/j.13652486.2012.02713.x
- Vincke, E., Boon, N., Verstraete, W., 2001. Analysis of the microbial communities on corroded
 concrete sewer pipes--a case study. Appl. Microbiol. Biotechnol. 57, 776–785.
- Vishwakarma, V., George, R.P., Ramachandran, D., Anandkumar, B., Mudali, U.K., 2014. Studies
 of detailed Biofilm characterization on fly ash concrete in comparison with normal and
 superplasticizer concrete in seawater environments. Environ. Technol. 35, 42–51.
 https://doi.org/10.1080/09593330.2013.808249
- Weilgart, L. s., 2007. The impacts of anthropogenic ocean noise on cetaceans and implications for
 management. Can. J. Zool. 85, 1091–1116. https://doi.org/10.1139/Z07-101
- Whitman, W.B., Coleman, D.C., Wiebe, W.J., 1998. Prokaryotes: The unseen majority. Proc. Natl.
 Acad. Sci. 95, 6578–6583. https://doi.org/10.1073/pnas.95.12.6578
- Witt, M.J., Sheehan, E.V., Bearhop, S., Broderick, A.C., Conley, D.C., Cotterell, S.P., Crow, E.,
 Grecian, W.J., Halsband, C., Hodgson, D.J., Hosegood, P., Inger, R., Miller, P.I., Sims,
 D.W., Thompson, R.C., Vanstaen, K., Votier, S.C., Attrill, M.J., Godley, B.J., 2012.
 Assessing wave energy effects on biodiversity: the Wave Hub experience. Philos. Trans. R.
 Soc. Math. Phys. Eng. Sci. 370, 502–529. https://doi.org/10.1098/rsta.2011.0265
- Wood, T.L., Guha, R., Tang, L., Geitner, M., Kumar, M., Wood, T.K., 2016. Living biofoulingresistant membranes as a model for the beneficial use of engineered biofilms. Proc. Natl.
 Acad. Sci. 113, E2802–E2811. https://doi.org/10.1073/pnas.1521731113

- Yañez, F., Gomez-Amoza, J.L., Magariños, B., Concheiro, A., Alvarez-Lorenzo, C., 2010.
 Hydrogels porosity and bacteria penetration: Where is the pore size threshold? J. Membr.
 Sci. 365, 248–255. https://doi.org/10.1016/j.memsci.2010.09.012
- Yip, T.L., Zhang, D.H., Chwang, A.T., 2002. Environmental And Safety Considerations For
 Design of a Perforated Seawall. OnePetro.
 https://onepetro.org/ISOPEIOPEC/proceedings/ISOPE02/All-ISOPE02/ISOPE-I-02 375/9056
- Yuan, H., Dangla, P., Chatellier, P., Chaussadent, T., 2015. Degradation modeling of concrete
 submitted to biogenic acid attack. Cem. Concr. Res. 70, 29–38.
- Zeppilli, D., Leduc, D., Fontanier, C., Fontaneto, D., Fuchs, S., Gooday, A.J., Goineau, A., Ingels,
 J., Ivanenko, V.N., Kristensen, R.M., Neves, R.C., Sanchez, N., Sandulli, R., Sarrazin, J.,
 Sørensen, M.V., Tasiemski, A., Vanreusel, A., Autret, M., Bourdonnay, L., Claireaux, M.,
 Coquillé, V., De Wever, L., Rachel, D., Marchant, J., Toomey, L., Fernandes, D., 2018.
 Characteristics of meiofauna in extreme marine ecosystems: a review. Mar. Biodivers. 48,
 35–71. https://doi.org/10.1007/s12526-017-0815-z
- Zobel, R.W., 1974. Control of morphogenesis in the ethylene-requiring tomato mutant,
 diageotropica. Can. J. Bot. 52, 735–741.
- 1230

1231